

Review

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The cancer stem cell: Evidence for its origin as an injured autoreactive T Cell

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Published: 14 February 2006

Received: 09 January 2006

Molecular Cancer 2006, **5**:6 doi:10.1186/1476-4598-5-6

Accepted: 14 February 2006

This article is available from: <http://www.molecular-cancer.com/content/5/1/6>

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Abstract

This review explores similarities between lymphocytes and cancer cells, and proposes a new model for the genesis of human cancer. We suggest that the development of cancer requires infection(s) during which antigenic determinants from pathogens mimicking self-antigens are co-presented to the immune system, leading to breaking T cell tolerance. Some level of autoimmunity is normal and necessary for effective pathogen eradication. However, autoreactive T cells must be eliminated by apoptosis when the immune response is terminated. Apoptosis can be deficient in the event of a weakened immune system, the causes of which are multifactorial. Some autoreactive T cells suffer genomic damage in this process, but manage to survive. The resulting cancer stem cell still retains some functions of an inflammatory T cell, so it seeks out sites of inflammation inside the body. Due to its defective constitutive production of inflammatory cytokines and other growth factors, a stroma is built at the site of inflammation similar to the temporary stroma built during wound healing. The cancer cells grow inside this stroma, forming a tumor that provides their vascular supply and protects them from cellular immune response.

As cancer stem cells have plasticity comparable to normal stem cells, interactions with surrounding normal tissues cause them to give rise to all the various types of cancers, resembling differentiated tissue types. Metastases form at an advanced stage of the disease, with the proliferation of sites of inflammation inside the body following a similar mechanism. Immunosuppressive cancer therapies inadvertently re-invigorate pathogenic microorganisms and parasitic infections common to cancer, leading to a vicious circle of infection, autoimmunity and malignancy that ultimately dooms cancer patients. Based on this new understanding, we recommend a systemic approach to the development of cancer therapies that supports rather than antagonizes the immune system.

Introduction

Understanding the pathomechanism of cancer is of primary interest in medical research. In the past century, several mechanisms were proposed: It was hypothesized that cancer arises out from a single cell that loses its differentiated state through sequential mutations [1]. This initiation-promotion-progression concept explains the steps in

a sequential process [2]. Later, this hypothesis led to the mutagenic and recently the oncogenic theories which hypothesize that defects in tumor suppressor genes are responsible for the development of cancer [3]. The impairment of cell-to-cell communication as a cause of cancer has also been postulated [4].

Mutations and other genetic abnormalities observed in cancer cells could also be caused by environmental effects, e.g., chemical carcinogens or life style factors such as alcohol or tobacco consumption or drug abuse [5]. The discovery of the cancer stem cell [6-8] lent support to the theory that cancer may develop out of a single cell, and raised the question of cancer stem cells arising from normal stem cells [9]. Indeed, if normal stem cells could undergo the type of mutations observed in tumor cells, this would potentially compromise the genetic stability of the organism. Therefore, the likelihood that normal stem cells are extremely well protected is demonstrated by their resistance to radiation and toxins [9].

One fascinating finding is that immunosuppressive cytotoxic antineoplastic therapies may on occasion cause the regression of a clinically established cancer. At first, applying this as a therapeutic strategy may seem counterintuitive, considering the fundamental role of the immune system in protecting the body against infectious organisms and aberrant cells. In addition, cancer itself is frequently immunosuppressive, so exacerbating a pre-existing immunosuppression may not seem like a rational strategy.

In this light, it appears paradoxical that the same degree of immunosuppression that is lethal in a bacterial or fungal infection actually benefits cancer suppression. In other words, the deletion of the T cell compartment that accompanies cytotoxic antineoplastic therapies [10] may facilitate cancer regression. This suggests that cancer itself may arise out of the immune system, potentially from the T cell compartment, which would explain why the suppression of cellular immunity could also lead to the suppression of the disease.

Another observation is that tumor cells are poorly immunogenic, despite the fact that tumor cells are antigenic [11,12]. Therefore, they do not generate a T cell-mediated immune response, and if so, it is of low intensity [13]. If tumor cells were derived from injured lymphocytes, particularly T cells that still share some functional properties with their normal counterparts, an immune tolerance to cancer cells could be explained, as the immune system is not made to attack itself. In pathological situations, T cells do attack self-tissue in a manner reminiscent of the autoreactive nature of cancer cells which have the ability to attack and invade host tissues. In other words, cancer cells behave like autoreactive lymphocytes. Here, we explore the evidence suggesting that such a mechanism could be at work during the development of cancer.

The prevalent genetic theories of cancer are built upon observations of genetic abnormalities in tumor cells. These theories do not generally take into account the dem-

onstrated importance of environmental factors in human cancer development. In a previous paper [14] we have shown that specific dietary deficiencies mimic the effects of chemical or radiation damage to DNA, which we propose plays an important role in human carcinogenesis and tumorigenesis. This observation allows us to consider cancer as a single disease, possibly developing from a single cancer stem cell. Based on this, we could assume that the observed genomic abnormalities in cancer cells are an effect rather than the cause of the disease. This idea also points to the direction of upstream events preceding the development of the malignant cell. We propose that identifying these events will be fundamental to understanding the pathomechanism of cancer. By exploring the functional similarities between lymphocytes and cancer cells, we provide an insight into this realm of possible upstream events.

The exterior cell surface layer (cell coat)

The lymphocyte cell coat is a labile structure, and the treatment of cells may lead to the loss of its components [15-20]. The cell coat plays an important role in lymphocyte functions including homing, cell mediated immunity, electrophoretic properties and antigen expression [21]; cell surface proteins are thought to be involved in cell propagation and differentiation [18]. After treatment with β -glucosidase [22], sialidase [23,24] and trypsin [25], lymphocytes lose their homing abilities. Cytotoxic lymphocytes transiently lose their cytotoxic ability after a brief papain treatment [26]. Lysis of the cell coat suppresses cell-mediated immunity [27-29]. Treatment by glycosidases including neuraminidase affects the bodily distribution of lymphocytes [23,24] and demonstrates alterations in their antigenicity [30-34]. Treatment with trypsin and neuraminidase reversibly eliminates the mitogenic response of lymphocytes [35,36]. The cell coat on thymocytes is significantly thicker than on splenic lymphocytes, [20] suggesting a role for the cell coat in T cell function. The cell coat of the lymphocyte cell membrane has been characterized using various stains [15-17], [37-39]. These investigations found high acid mucopolysaccharide content with a significant number of acidic amino sugar end groups.

Cancer cells also exhibit an exterior cell surface coat [40-45]. The similarities between the cell coat of normal and leukemic lymphocytes have been investigated [39,41]. Pathological lymphocytes (CLL) have a uniformity of staining similar to their normal counterparts, with some differences observed with cationic stains that could be due to a decrease in the sialoprotein of the cell coat of CLL cells. With some similarity to lymphocytes, the tumor cell coat has been suggested to play a role in cell contact and adhesion, cell recognition [44], as well as the capacity to metastasize [46].

The tumor cell coat is also sensitive to neuraminidase [47-49] and can rapidly re-grow following treatment with the enzyme [50]. The enzyme treatment also changes the immunological properties of tumor cells. Trypsin and EDTA removes the tumor cell coat [51]. The cell coat is involved in the mechanism by which tumor cells escape cellular immune attack [45,52-54]. The degradation of the cell coat by brief hyaluronidase treatment of glioma cells sensitizes them to cytotoxic lymphocyte attack [52,53]. Although normal human glial cells also produce hyaluronic acid, glioma lines produced significantly more. Hyaluronidase-sensitive coats have been found on a variety of murine sarcoma and carcinoma cell lines [54]. It appears that a mucopolysaccharide coat on tumor cells impedes the successful use of immunotherapy. It was demonstrated that the displacement of the tumor cell coat by charge-functionalized lipids or polycationic substances leads to tumor cell apoptosis and tumor destruction [45,55,56].

It is demonstrated that the cell coat of lymphocytes and tumor cells are functionally significant. The degradation/removal of cell coat significantly impacts the functionality of both tumor cells and lymphocytes; therefore, tumor cell isolation methods could alter the functionality of isolated cells. In other words, with the loss of the cell coat, lymphocytes lose fundamental functions, i.e., cannot attack target cells, while tumor cells also lose cell contact and adhesive properties, as well as the ability to metastasize. In addition, tumor cells become sensitive to apoptosis.

Activation of coagulation

The activation of coagulation occurs during tissue injury as well as in various pathologies. Infection leads to an inflammatory reaction as well as the activation of coagulation, as there is a crosstalk between these functions [57-59]. Blood coagulation components can inhibit or amplify the inflammatory response. Blood clotting is initiated when pathogenic components such as endotoxin or inflammatory cytokines induce the synthesis of tissue factor on leukocytes [60]. The coagulation cascade is subsequently triggered. The formation of negatively charged membrane phospholipid surfaces amplifies the coagulation reaction [61]. Natural anticoagulant pathways such as the protein C anticoagulant pathway limit the coagulation process, thereby suppressing the inflammatory response including reducing inflammatory cytokine secretion [62], decreasing NF- κ B signaling [63], minimizing leukocyte chemotaxis [64] and endothelial cell interactions [65], and suppressing apoptosis [66].

Platelets are also involved in the link between inflammation and coagulation. Inflammatory cytokines such as IL-6 or IL-8 increase platelet production, and such platelets

are more thrombogenic [67]. In addition, the platelets release the CD40L protein, a potent proinflammatory mediator, which subsequently induces tissue factor synthesis [68,69] and amplifies the secretion of proinflammatory cytokines [70,71]. This in turn leads to a progressive cycle that ultimately can produce severe vascular and organ injury.

In 1865, Trousseau first described a cancer-associated condition now called migratory thrombophlebitis in which a spontaneous coagulation of the blood occurs in the absence of inflammatory reactions [72]. It manifests as migratory thrombosis in the superficial veins of the chest wall and arms, but it can occur in other sites as well. This condition is a variant of venous thromboembolism. Thrombosis is a frequent complication of malignancy, and thromboembolic death is the second leading cause of mortality in cancer [73,74]. Malignant cells interact with the blood coagulation system by releasing procoagulant and fibrinolytic substances and inflammatory cytokines [75-85]. In addition, direct interaction with endothelial cells, monocytes/macrophages, and platelets also leads to localized clotting activation [85-87]. Similar to normal activated inflammatory cells, malignant cells release tissue factor [75-77] which promotes the formation of fibrin deposits in the tumor cell microenvironment [88-90].

The fibrin gel matrix along with other connective tissue components form the basis for the tumor stroma, a matrix in which tumor cells are dispersed and which provides the vascular supply as well as a barrier against rejection by the cellular immune system [89]. The tumor stroma shares properties in common with the temporary stroma of a healing wound [91]. Similar to the fibrin coating on macrophages [92], the observed fibrin coating of tumor cells is involved in the mechanism by which tumor cells escape destruction by NK cells [93,94]. Histological evidence suggests that inflammatory lymphocytes are confined to the tumor-host interface, and do not significantly penetrate the tumor [89,95]. Malignant cells secrete inflammatory cytokines such as TNF- α and IL-1 β that downregulate the anticoagulant system of vascular endothelial cells [96,97]. The secretion of IL-8 promotes new blood vessel formation, [98] and the fibrin deposited around tumor cells facilitates angiogenesis [99-101].

Tumor cells attach to the vascular endothelium and promote the adhesion of leukocytes and platelets [102-105]. Monocytes and macrophages also home in on vascular surfaces due to inflammatory stimuli [106-108]. In response to inflammatory molecules, complement, lymphokines and immune complexes, these cells subsequently secrete procoagulant tissue factor; tumor-associated macrophages express significantly higher levels of tissue factor than control cells [109,110]. These macro-

phages also increase their fibrinolytic enzyme production [111].

Both human and animal cancer causes platelet aggregation *in vitro* and *in vivo* [112-114]. The ability of tumor cells to aggregate platelets and secrete plasminogen activator correlates with their metastatic potential [115]. Indeed, thrombocytopenia reduces the metastases of tumors [116,117] as do compounds capable of reducing platelet aggregation [117-125]. These include aspirin, prostaglandins and other nonsteroidal (NSAID) anti-inflammatory drugs. A reduced risk of fatal colon cancer has been observed among aspirin users [120-122]. Administration of heparin and fibrinolytic also reduces the incidence of experimental metastases [126-128], while the administration of anti-fibrinolytic agents increases their incidence [129,130].

Cancer treatment by surgery, cytotoxic antineoplastic drugs and hormonal therapy all contribute to the hypercoagulable state and risk factors for thromboembolism in cancer patients [131,132]. The risk of fatal pulmonary embolism increases four-fold after surgery in cancer patients [133,134]. Chemotherapy drugs including cisplatin, mytomicin C and tamoxifen as well as high-dose and multi-drug regimes increase the risk of thrombotic complications [135-139]. Prophylactic treatment with warfarin reduces this risk (140). The use of hematopoietic growth factors subsequent to chemotherapy was shown to induce thrombosis in breast cancer patients [141,142]. Venous thrombosis could also be a marker for an otherwise asymptomatic cancer [143,144].

Similarly to a normal inflammatory reaction, activation of coagulation takes place in cancer. The events of tumor stroma development are comparable to wound healing [91] and it is possible that tumor formation may be associated with defective wound healing initiated by an inflammatory reaction due to infection and/or tissue injury. Therefore, we believe it is important to investigate potential links between infection, inflammation and cellular immune response in searching for the origins of the cancer cell.

Infection and inflammation

The etiological role of infectious agents has been indicated in various cancers. In 100 cases of human leukemia, *Mycoplasma*, *Salmonella*, *Micropolyspora*, *Mycobacterium*, *Absidia*, pseudorabies virus and adenovirus antigens were commonly detected in the patient's sera [145]. Hepatotropic viruses (hepatitis B and C) cause hepatic necrosis followed by hepatocellular, B cell and gastric malignancies [146-149]. Antiviral therapy of hepatitis C infection led to the regression of virus-associated B cell lymphoma [150]. Adenoviral infection has been associated with

childhood leukemia [151] and cytomegalovirus infection with testicular cancer [152]. *Helicobacter pylori* infection is widespread in the population (an estimated 40–80% infected) and is linked to gastric cancer and mucosa-associated lymphoid tissue (MALT) lymphoma [153,154]. A reversal of lymphoma-induced neutropenia has been observed with the eradication of *H. pylori* infection [154]. Simian virus 40 (SV40) is associated with human brain cancers and non-Hodgkin's lymphoma [155]. Ocular adnexal lymphoma is linked to *Chlamydia psittaci* infection, and the reversal of lymphoma was observed with pathogen-eradicating antibiotic therapy [156]. The list continues: Cervical intraepithelial neoplasia (CIN) is associated with human papilloma virus (HPV) infection with a co-etiological presence of chronic bacterial cervicitis [157-159]. *Mycoplasma* and HPV association was found to be dominating. The role of mycoplasma in the dysplasia of the uterine cervix and development of CIN has also been demonstrated [160].

Mycoplasmas are particularly interesting due to their widespread presence in the human population. Although many mycoplasmas are not directly pathogenic in humans, they are associated with many diseases [161-165]. Mycoplasmas have co-leukemogenic activity [166-168] and are found to increase tumor cell invasiveness [169]. In approximately half of the examined cases, mycoplasma DNA was present in ovarian and gastric carcinoma specimens [170,171]. In gastric, lung, esophageal, breast and colon cancers as well as glioma specimens, *Mycoplasma hyorhinis* was detected in about 50% of the cases [172]. Mycoplasmas are known to cause chromosomal changes [173]. Mixed *Mycoplasma pneumoniae* and influenza virus infection induced lung cancer in an animal model [174]. The direct role of the AIDS-associated *Mycoplasma fermentans* and *Mycoplasma penetrans* in oncogenesis has been investigated [175]. These mycoplasma strains induced gradual malignant transformations that eventually became irreversible. Besides its direct oncogenic potential, *Mycoplasma fermentans* was found to exhibit a unique cytotoxic effect on the undifferentiated myelomonocytic lineage, but not on differentiated myelomonocytic cells [176]. The depletion of immature myelomonocytic cells likely contributes to the functional immunodeficiency present in cancer patients.

In response to pathogens, the host mounts a protective inflammatory response. Immune cells migrate to the area of infection and produce inflammatory messengers called cytokines. Initially, cells of the innate immune system (macrophages, neutrophils, NK cells) become involved, followed by the activation of cells of the adaptive immune system. These include antigen-presenting cells (APCs), T and B cells, which play an important role in propagating the inflammatory response. T cell inflammation plays a

major role in antitumor immune responses. Key regulators of T cell-mediated response are the T helper (Th) cells that secrete the cytokines orchestrating this response. The two subtypes Th1 and Th2 cells produce cytokines stimulating cellular and humoral immune responses.

Intracellular pathogens (e.g., viruses, mycoplasmas) use the Toll-like receptor (TLR) signaling mechanism to escape host defenses [177]. Pathogen-associated molecular patterns on the surface of mycoplasmas engage TLRs 1, 2, and 6 on the surface of APCs that lead to a Th2-type polarization of the immune response and the secretion of IL-10, IL-4, IL-5 and IL-13 [178-180]. These cytokines are antagonistic to Th1 type cytokines (TNF- α , IL-2, IFN- γ , IL-6, IL-12); excessive production of either type of cytokine upsets the homeostatic balance needed to maintain a proper mix of cellular and humoral immune responses. Utilizing this mechanism, mycoplasmas suppress cell-mediated immunity, which allows them to persist and predispose the host for colonization by other pathogens. The observation that leukemia patients were colonized by over half a dozen pathogens besides mycoplasmas [145] suggests that suppression of the cellular immune system provides a fertile ground for a variety of pathologies.

Besides regulating innate and adaptive immune responses, cytokines are involved in cell growth and differentiation. Normally, the secretion of cytokines is of short radius and limited duration, typically regulating self or adjacent cell functions. The activity of cytokines is tightly regulated, and there is evidence that cytokines contribute to inflammatory autoimmune diseases [181-184] and malignancies. Similarly to activated T cells, various tumor cells secrete immune response-polarizing cytokines (IL-10, IL-6, IL-8, IL-13, TGF- β) serving as autocrine and/or paracrine growth factors for the cancer [185-199]. The progression of the disease and patient survival was correlated with increasing levels of cytokine secretion [200]. This secretion is frequently constitutive, leading to elevated serum levels of cytokines in malignancies including melanoma, non-small cell lung carcinoma, renal cell carcinoma and bladder carcinoma [186-190,201]. In addition, tumor cells can induce IL-10 in the tumor environment [191]. IL-10, the most potent Th2 polarizing cytokine, suppresses the tumoricidal activity of macrophages [202], blocks presentation of tumor antigens to professional APCs [203-205], and inhibits tumor-specific cytotoxic T cells [206]. However, in cancers both cellular and humoral immune response may be depressed, as in the absence of IL-4 production IL-10 secretion alone cannot induce a Th2-type response.

It appears that the immune response becomes distorted at multiple levels during the development of cancer. First, infectious agents may act in concert to subvert cellular

immunity, thereby upsetting the homeostatic balance of a proper mix of cellular and humoral immune response. This leads to an aberrant cytokine-signaling that results in depressed apoptosis and excessive proliferation [207,208]. Cytokines seem to be the key substance of apoptosis of leukemic cells [207]. Abnormal inflammatory cytokine secretion by tumor cells reinforces the existing imbalances and thus promotes disease progression. Similarly to T cells, cancer cells use inflammatory cytokines as autocrine and paracrine growth factors, suggesting a functional relationship between cancer cells and cells of the immune system.

Infection, autoimmunity and cancer

Several lines of evidence suggest a direct relationship between infection, autoimmunity and cancer. Hepatitis B and C viruses are involved in an autoimmune condition that precedes the development of hepatocellular carcinoma [209]. Data also demonstrate a higher prevalence of B-cell non-Hodgkin's lymphoma in HCV-infected patients with autoimmune manifestations [147-149] including Sjorgren syndrome [210], cryoglobulinemia [211,212] and systemic lupus erythematosus (SLE) [213,214]. Adenovirus infection is associated with childhood leukemia, (151) and family studies in acute childhood leukemia have shown possible associations with autoimmune disease [215]. Epstein-Barr virus [216] and human T lymphotropic virus type 1 infection [217] is associated with abnormal lymphoproliferation and Hodgkin's lymphoma. Cytomegalovirus infection is linked to autoimmunity [218] and testicular cancer [152].

H. pylori infection can lead to autoimmune neutropenia and MALT-lymphoma [154] in addition to its well-established role in the development of gastric cancer. Systemic rheumatic disease has also been linked to lymphoid malignancy [219]. These findings underline a close relationship between infection, autoimmunity and proliferative disorders, possibly mediated by an abnormally functioning cytokine signaling network [220].

Antinuclear antibodies (ANA) were demonstrated in the sera of 19% of patients with malignancies in the absence of overt autoimmune manifestations [221]. In cancer patients, a large number of autoantibodies are observed against tissue-specific antigens, nucleoproteins, membrane receptors, proliferation-associated antigens, tissue-restricted antigens, etc. [reviewed in [222]]. Autoimmune connective tissue disorders are also commonly associated with malignancies [223]. It was reported that gastric atrophy and pernicious anemia carries a risk for gastric carcinoma 18 times that of the population average [224]. It appears that a variety of infections may induce autoimmune serological features without overt autoimmune disease or organ involvement [225]; however, this condition

may progress to clinical autoimmune disease and malignancy if impaired T cell function prevails. Such condition develops at a higher frequency among the elderly [226].

It was observed 30 years ago that a low percentage of human T cells (3.4%) have the ability to form auto-rosettes with autologous erythrocytes; in breast cancer and melanoma patients, the ratio was elevated to 6.1% and 7.4%, respectively [227]. This observation implied that some level of autoreactivity is normal, confirmed later by studies on T cell tolerance [228,229]. However, the observation also pointed to an elevated level of autoreactive T cells involved in cancer. The mechanism of activation of an autoreactive T cell response was linked subsequently to bacterial and viral infections through the process of molecular mimicry [218,230-234] in which pathogen-derived peptides mimic self-peptides. This phenomenon was studied in animal models [235-240] and was supported by clinical observations [241-243]. As a highlight, when lymphocytic choriomeningitis virus (LCV) antigens were expressed in the pancreas of transgenic mice, infection with the virus led to autoimmunity and diabetes [239].

H. pylori antigens mimic epitopes on H⁺, K⁺-adenosine triphosphatase in the gastric mucosa [230] thereby activating cross-reactive gastric T cells. Viral peptides mimic sequences on myelin basic protein [234], leading to multiple sclerosis. Cytochrome c (cyt c) as an antigen was used to study how self-proteins prime autoreactive T cell responses [244,245], as SLE patients possess autoantibodies to cyt c [246]. When non-self cyt c was co-administered with the self-protein, B cells specific for the foreign antigen primed autoreactive T cells that led to breaking tolerance to self-cyt c. The same autoimmune phenomenon occurs in the LCV transgenic mice when LCV antigens on pancreatic cells and the intact virus antigens are co-presented to the immune system [239]. Therefore, it is quite likely that autoimmunity spontaneously develops during a variety of infections when antigens on microorganisms mimic self antigens and are presented together, breaking T cell tolerance.

The presence of autoreactive T cells has been observed in healthy persons, which indicates a role for these cells in immune defense. If autoreactive T cells were always absent from the T cell repertoire, the responsiveness toward foreign antigens that resemble self-antigens would be reduced. This notion is supported by the observation that T cells which recognized variants of self-antigen are of lower avidity than those recognizing a foreign antigen [247,248]. Also, tolerance to self-antigen reduced T cell variants for these peptides as well as the diversity of T cell receptor α and β -chain sequences of self-specific T cells [249,250]. It appears that some level of autoreactive T

cells is necessary for immune defenses. Clinical autoimmunity may develop when persistent infection provides a continuing high dose of antigenic stimulus, [251] and this situation could predispose patients for the development of proliferative disorders.

Defective apoptosis

Normal tissue development requires damaged, dangerous or unnecessary cells to be eliminated while healthy cells survive. The survival of harmful or damaged cells can lead to various pathologies. The evolutionarily conserved mechanism of apoptosis eliminates unwanted or abnormal cell populations. Lymphocytes require IL-2, IL-4, IL-7, IL-9 and IL-15 for viability [252,253], and withdrawal of these cytokines leads to apoptotic cell death. Leukemia patients who went into complete remission following chemotherapy developed a different type of leukemia after being placed on IL-2 therapy [185]. IL-2 is an essential cytokine for the viability of activated T-cells [254], suggesting a link between the survival of activated T-cells and leukemic cells. Myeloid leukemia cells are also cytokine-dependent and undergo apoptotic cell death following cytokine withdrawal [253]. The various immune response-polarizing cytokines that tumor cells secrete [185-201] inhibit chemotherapy- or radiation-induced apoptosis [256-261]. There are myeloid leukemia cell lines that have become independent of an external cytokine supply [257], but generally cytokines can protect both normal and cancer cells against apoptosis induced by various cytotoxic agents. The persistence of infectious agents and chronic inflammation in cancer patients promotes NF- κ B activation and inflammatory cytokine production, thereby contributing to the diminished apoptosis of abnormal cells [262,263].

The completion of immune response against pathogenic microorganisms requires the deletion of activated T and B cells that participated in the immune defenses, particularly self-reactive ones [264] (although a fraction of them survive as memory cells). Apoptosis plays an important role in the regulation of peripheral immunity through the Fas/APO-1 cytotoxic pathway. Defective apoptosis can lead to autoimmune disease [265,266] and cancer [267,268]. As cancer cells are not immortal, they maintain a program for apoptotic cell death [269].

The apoptosis marker Fas receptor (FasR) is expressed on numerous cell types, whereas the Fas ligand (FasL) is mainly expressed on T cells [266]. FasL mediates the apoptosis of effector T cells as part of an immune response termination and tolerance development. FasL is also expressed in "immune-privileged" tissues such as the brain, testes and eyes with the purpose of preventing inflammation. Mutations in Fas or FasL can lead to autoimmune disease [270,271]. Similarly to cytotoxic T

cells, various tumor cells also express FasL and use it to induce apoptosis of invading lymphocytes. Breast tumor cells express FasL that can kill Fas-sensitive lymphoid cells [272]. The co-expression of Fas and FasL was observed in brain tumors that can use this mechanism to obtain a proliferating advantage by "counter-attacking" tumor-infiltrating activated Fas-sensitive T lymphocytes [273,274]. Similar observations have been made in Ewing sarcoma [275], gastric cancer [276], cholangiocarcinoma [277], B cell chronic lymphocytic leukemia (B-CLL) [278], colon adenocarcinoma [279-281], head and neck cancer [282], lung carcinoma [283], esophageal carcinoma [284], ovarian carcinoma [285], lymphoma [286], pancreatic carcinoma [287], melanoma [288], and other malignancies [289,290]. Childhood glial tumor cells (but not normal cells) in the brain express the common leukocyte-associated antigen and Fas [273].

The expression of apoptosis-related molecules on the surface of both neoplastic cells and cytotoxic lymphocytes (CTL) in tumor specimens raises the question of whether neoplastic cells are formed from CTLs by a premature termination of the apoptotic mechanism. Indeed, neoplastic cells behave like CTLs in their expression of FasL and in the induction of apoptotic death of activated T cells, as well as other cancer cells carrying a functional FasR [291,292]. In other words, cancer cells continue to act like T cells performing their immune-regulating functions.

Discussion and therapeutic implications

Infections by various pathogenic microorganisms are a common occurrence in humans and other animals. In response to invading pathogen(s), an inflammatory reaction develops in the host organism. Initially, the innate immune system becomes involved, followed by the development of an adaptive immune response. Activated leukocytes produce inflammatory cytokines and chemokines as well as other growth factors aimed at clearing up the infection and facilitating tissue healing. The inflammatory reaction at the infection site triggers a variety of physiological responses. Antigen-presenting cells activate T and B cells in response to molecular patterns expressed on the surfaces of pathogenic microorganisms. Intracellular pathogens are overcome by the cellular immune response; in addition, the T cell inflammatory reaction is also key to antitumor immunity. Activated T helper 1 (Th1) cells secrete specific cytokines orchestrating this response.

Pathogenic microorganisms, however, have evolved strategies to evade immune surveillance in order to persist in the host. Several intracellular pathogens including mycoplasmas and viruses deploy molecular patterns on their surfaces that trigger a Th2-type (humoral) immune response and consequently depress cellular immunity. In addition, some infections such as the mycoplasmas

remain sub-clinical, and by subverting the cellular immune response, these microorganisms predispose the host for colonization by other pathogens eventually leading to various pathologies.

Molecular mimicry is initiated when viruses integrate host genes within their genome, [293] and pathogens with host-like genes may have a survival advantage over those lacking such traits. Animal viruses are capable of fusing with prokaryotic cells that may facilitate gene transfer between distant microbial taxa [294]. Influenza virus hemagglutinin A sequences have been located in the p37 protein of *Mycoplasma hyorhinis*, and this protein increases tumor cell invasiveness [295]. The exchange of genes among various microorganisms [296] leads to the development of antibiotic resistance. Gene uptake also occurs by phagocytosis of apoptotic bodies [297,298] while High Mobility Group (HMG) proteins, commonly associated with human DNA, may facilitate this process in bacteria [299].

When antigens from pathogens mimic self-antigens in the process of molecular mimicry, cross-reactive T cells may be generated. The study on breaking T cell tolerance with co-administered foreign and self-cytochrome c is a sobering reminder of just how easy is to induce autoimmunity. However, evidence also demonstrates that a low level of autoimmunity is normal and necessary to mount an effective immune response to infections. Clinical autoimmunity may develop if a continuing high-dose antigenic stimulus persists, as in cases of chronic infection. In addition, there is also evidence that autoimmunity can lead to proliferative disorders.

As discussed, normal tissue development requires the elimination of dangerous and abnormal cells, and autoimmune T cells belong into this category. With the completion of the immune response, the evolutionarily conserved mechanism of apoptosis eliminates effector T cells, leading to immune response termination and tolerance development. However, defective apoptosis can lead to autoimmunity and cancer.

We propose that an aberration in the apoptosis process leads to formation of the cancer stem cell from autoreactive T cells. In support of this observation, *Helicobacter*-induced gastric epithelial carcinoma was found to originate from bone marrow-derived cells [300]. This is direct proof of cancer that is not arising from mutated epithelial cells. Also, the cytotoxic T lymphocyte-associated antigen-4 (CTLA-4), a regulator of the effector function of T cells, is expressed in various leukemias and solid tumors [304]. This suggests a link between CTLs, hematopoietic neoplasias and solid tumors.

Further evidence: the common acute lymphoblastic leukemia antigen was detected on glioma [301] and melanoma [302] cell lines. The melanoma-associated PRAME antigen is expressed both in leukemias and some solid tumors [303]. The majority of leukemia and lymphoma cells test positive for the leukocyte common antigen (CD45) [305]. Seminoma [306], rhabdomyosarcoma [307] and some metastatic undifferentiated and neuroendocrine carcinomas [308] have also been found to express CD45. The myeloid antigen Leu-7, typically expressed on natural killer (NK) cells and T cell subsets, was detected on small cell lung carcinoma [309,310] and a variety of other solid tumors including astrocytoma, neuroblastoma, retinoblastoma, carcinoid tumors, etc. [311]. Neoplastic cells of Hodgkin's disease expressing Leu-7 may be related to NK cells or T cells rather than B cells [312]. We propose that the unexpected presence of some T cell markers on cancer cells may provide an insight into their origins. In addition, the observation that cancer stem cells embedded in an environment of normal host tissue can undergo a differentiation process (during which surface markers of lymphoid origin disappear) explains the absence of leukocyte-derived surface antigens in some solid tumors.

In benign colonic adenomatous polyps and synchronous adenocarcinoma, comparable and very large numbers of genomic alterations (>10,000 events per cell) were found [313], demonstrating massive genomic damage characteristic of apoptosis as opposed to sequential mutations. In addition, this demonstrates that genomic instability precedes the development of a malignant state, indicating that malignancy is an effect rather than the cause of genetic abnormalities in cancer cells. It is therefore reasonable to conclude that there is no fundamental difference between benign and malignant tumors, and that possibly just a small difference in the dysregulation of proliferative controls leads to a malignant phenotype.

We further propose that the resultant cancer stem cell still preserves some functions of an effector T cell, such as homing in to sites of inflammation such as the inflamed bronchi of a cigarette smoker, the damaged liver of an alcohol abuser, an *H. pylori*-infected gastric mucosa, an HPV-infected uterine cervix, an inflamed colon, etc. The cancer cell retains some capabilities of an effector T cell to secrete inflammatory cytokines (even if in an aberrant, constitutive fashion), thereby distorting local immune responses, disabling cytotoxic T cells and diminishing apoptosis in its environment.

Like normal activated inflammatory cells, cancer cells activate the coagulation system, leading to the formation of the tumor stroma in which tumor cells proliferate. Dvorak in his paper entitled "Tumors: wounds that do not heal" [91] succinctly described similarities between the forma-

tion of the temporary stroma of a healing wound and tumor stroma development. While the cancer cell continues to act as if it participated in a wound healing process, it actually enlarges the wound stroma due to its constitutive secretion of tissue factor, inflammatory cytokines and other growth factors which also provide stimuli for the propagation of the malignant cells. This leads to an ever-continuing cycle of tumor growth.

Every human cell has the ability to repair itself, and cancer cells retain some of this capacity [314]. As cancer stem cells exhibit plasticity similar to normal stem cells, we propose that a cell-to-cell communication between cancer stem cells and surrounding host tissues allows tumor cells to develop varying degrees of differentiated phenotypes resembling cells of normal differentiated tissues. This in turn leads to the emergence of various tumor types and creates the illusion of a great multitudes of cancers.

It has been long known that cancer cells, besides growing inside tumors, also circulate in the blood [315-317]. This is easy to rationalize if cancer cells are indeed damaged autoreactive T cells, and also provides an explanation for metastasis formation. Cancer cells interact with neutrophils, macrophages and platelets that lead to the formation of micrometastases that can remain in the blood for a long time [318]. These aggregates persist even after adjuvant chemotherapy, although in reduced numbers. Larger cell clumps are more effective in promoting metastases than smaller ones [319]. With the progression of inflammation in cancer patients, the circulating micrometastases find new sites of proliferation that lead to the formation of metastases.

Current cancer therapies are tumor-centric, as tumors are equated with cancerous disease. Main therapeutic modalities include the surgical removal of tumors as well as radiation and chemotherapies. All of these contribute to the hypercoagulable state and risk of thromboembolism, which have a significant negative impact on the morbidity and mortality of cancer patients. If tumor cells did originate from T cells, any therapeutic approach targeting tumor cells will likely diminish T cell function. Cytotoxic antineoplastic therapy represents an extreme situation in this regard, resulting in the deletion of even resting T cells, the reconstitution of which takes several months [10]. This makes the combination of chemotherapy and immunotherapy an unrealistic proposition.

If cancer cells indeed originate from damaged autoreactive T cells, our current views on cancer immunotherapy need to be revised. The immune system was not made to attack itself, and this is supported by the unresponsiveness of the cellular immune system to cancer even if tumor cells are antigenic. When we attempt to induce an immune

response against tumors, we run the risk of developing autoimmune disease [320] and ultimately, secondary malignancies.

The suppression of the immune system by chemotherapeutic agents and radiation encourages the propagation of microbial and parasitic infections already present in cancer patients. However, pathogenic microorganisms are intimately involved as co-etiological agents in the development of various malignancies via molecular mimicry-induced autoimmunity, and maintain a cytokine milieu that favors proliferation as opposed to apoptosis. Current immunosuppressive cancer therapies establish the conditions for disease recurrence as well as the emergence of new primary tumors, which is in fact, a common experience. Also, the cancer patient's system appears to retain a "memory" of the disease as the risk of developing another cancer is higher than those who have never had the disease. This memory could be attributed to autoimmune memory T cells, reactivated by recurrent infections which become cancerous later on as a consequence of defective apoptosis.

The eradication of pathogens could have a favorable effect on the course of malignant diseases, as demonstrated by therapies of HCV [150], *H. pylori* [154], and *Chlamydia psittaci* infections [156]. Mycoplasmas are difficult to eradicate and require high-dose, long-term antibiotic therapies, but even after that the pathogens are found to persist [321]. There are no therapies for many viral infections at this time. With our new understanding of the mechanism of TLR signaling, opportunities have opened for overcoming these types of pathogens. Very recently, a therapeutic oral mycoplasma vaccine was described [322], the principle of which could be utilized for the therapy of other intracellular infections.

If defective apoptosis of autoreactive T cells leads to the emergence of the cancer stem cell, our research must focus on the physiological events associated with apoptosis. Any therapeutic approach downstream from this step is merely symptomatic, and offers little hope of defeating cancer. A century of accumulated evidence on the use of immunosuppressive cancer therapies supports this observation.

It was demonstrated that the exterior mucopolysaccharide cell surface coat on cancer cells protects them from apoptosis [52,53]. Kovacs has explored this understanding to the greatest degree by synthesizing unsaturated aminolipids capable of displacing the cell coat on tumor cells [45]. Administration of these compounds led to the apoptotic death of a variety of tumor cells *in vitro* and *in vivo* [45]. Normal lymphocytes are less sensitive to the apoptotic effects of a fatty acid mixture than leukemic cells,

although they do show some sensitivity [323]. This observation may explain why the continuing administration of synthetic unsaturated aminolipids led to a diminishing efficacy of the therapy [324], as normal lymphocytes are also surrounded by an exterior cell surface layer coat essential for their functions.

Endocrine hormonal signaling also affects apoptosis. Corticosteroids facilitate the apoptosis of lymphocytes and exert an immunosuppressive effect when the organism is subject to prolonged stress. Stress also down-regulates the digestive functions of the gut, including those of the stomach and pancreas. This in turn suppresses the uptake of critical nutrients that are essential for genomic stability [14]. It was reported that breast cancer patients as a group exhibit a depressed thyroid function [14], suggesting an etiological role for thyroid deficiency in neoplasia. Thyroid function is profoundly affected by the iodine supply, and thyroid, breast and gastric cancers have been linked to iodine deficiency [14]. Previously we have pointed out that critical nutrient deficiencies mimic the effects of chemical or radiation damage to DNA, and suggested that the correction of these deficiencies could reverse the progression of malignant proliferation [14].

In the past century, insufficient attention was paid to the role of dietary factors in the development and progression of malignant diseases. No Recommended Daily Allowances (RDAs) are available for a number of essential nutrients, and where available, the RDA is of questionable value. Iodine, a vital micronutrient, is an example: the current WHO recommendation for iodine is 0.15 mg/day. However, some Japanese consume as much as 50–80 mg of iodine/day through their seaweed rich diet [325] and exhibit significantly lower rates of the major cancer types than seen in the Western world [14]. In addition, iodine supplementation clinical trials have demonstrated that an iodine intake vastly exceeding the RDA (more than 6,000 times higher) was both safe and clinically useful [326,327]. This could not possibly be the case if the RDA for iodine had been correctly determined. Similar clinical observations were made for high-dose administration of folate and vitamin B₁₂ [328,329] as well as vitamin C [330]. These findings question the accuracy of dietary RDAs, and suggest that current regulatory initiatives aimed at restricting the active ingredient contents in vitamin supplements are based on an erroneous scientific rationale.

It is also important to recognize that vitamin and mineral levels have significantly declined over the past 60 years in our food supply [reviewed in [331]] possibly due to intensive agricultural production methods and industrial food processing. Experience teaches us that in the Western world, despite an abundance of food, people have diffi-

culties in meeting their nutritional needs, demonstrated by now-rampant obesity as well as the historically proven explosion of degenerative diseases including cardiovascular diseases, diabetes and cancer. This suggests that we are still far from understanding the dietary needs of the human organism.

It is known that diabetics develop malignancies at a higher frequency than the population average [332,333], which implicates pancreas dysfunction in the etiology of cancer. Besides secreting digestive enzymes, the pancreas is also a source of hormonal regulators. We hypothesize that a combined effect of adrenal, thyroid and pancreas dysfunction may predispose patients for neoplasia in a process promoted by dietary deficiencies as well as life-style factors including prolonged stress, poor hygiene, smoking, alcoholism and drug abuse, all of which are known to subvert immunity. It appears that we need to make the most important scientific discoveries in the simplest things, i.e., how to conduct our lives in a manner optimal for well-being. Therefore, the main operative principle of health care should be prevention.

To finally defeat cancer, our research need to focus on the identification of those endocrine-signaling mechanisms that enable CTLs to complete their mission of apoptotic elimination of autoreactive T cells. We must abandon our focus on the tumor cell as far as the development of cancer therapeutics are concerned, as the destruction of cancer itself negatively impacts the immune system, thereby reactivating the vicious circle of infection, autoimmunity and malignancy that ultimately dooms cancer patients. By redirecting our focus toward physiological events preceding the formation of the cancer stem cell, we will be able to overcome this scourge that has haunted humanity since time immemorial. A systemic approach described in a previous paper [14] offers an alternative to current cancer therapies that works with the immune system, and which helps to re-establish homeostatic balance in the human body.

References

- Sell S: **Cellular origin of cancer: de-differentiation or stem cell maturation arrest.** *Environ Health Perspect* 1993, **101(Suppl 5)**:15-26.
- Pitot HC, Goldsworthy T, Moran S: **The natural history of carcinogenesis: implications of experimental carcinogenesis in the genesis of human cancer.** *J Supramol Struct Cell Biochem* 1981, **17**:133-146.
- Trosko JE, Chang CC, Madhukar BV, Dupont E: **Oncogenes, tumor suppressor genes and intercellular communication in the 'oncogeny as partially blocked ontogeny' hypothesis.** In *New Frontiers in Cancer Causation* Edited by: Iversen OH, Wash DC. Taylor and Francis Publishers; 1993:181-197.
- Trosko JE, Chang CC, Medcalf A: **Mechanisms of tumor: potential role of intercellular communication.** *Cancer Invest* 1983, **1**:511-526.
- Wogan GN, Hecht SS, Felton JS, Conney AH, Loeb LA: **Environmental and chemical carcinogenesis.** *Semin Cancer Biol* 2004, **14**:473-486.
- Passegue E, Jamieson CH, Ailles LE, Weissman IL: **Normal and leukemic hematopoiesis: are leukemias a stem cell disorder or a reacquisition of stem cell characteristics?** *Proc Natl Acad Sci USA* 2003, **100(Suppl 1)**:11842-11849.
- Singh SK, Clarke ID, Terasaki M, Bonn VE, Hawkins C, Squire J, Dirks PB: **Identification of a cancer stem cell in human brain tumors.** *Cancer Res* 2003, **63**:5821-5828.
- Dick JE: **Breast cancer stem cells revealed.** *Proc Natl Acad Sci USA* 2003, **100**:3547-3549.
- Dean M, Fojo T, Bates S: **Tumour stem cells and drug resistance.** *Nature Rev Cancer* 2005, **5**:275-284.
- Mackall CL: **T-cell immunodeficiency following cytotoxic anti-neoplastic therapy: A review.** *Stem Cells* 2000, **18**:10-18.
- Boon T, Coullie PG, Van den Eynde B: **Tumor antigens recognized by T cells.** *Immunol Today* 1997, **18**:267-268.
- Boon T, Old LJ: **Cancer tumor antigens.** *Curr Opin Immunol* 1997, **9**:681-683.
- Melief CJ, Toes RE, Medema JP, Van der Burg SH, Ossendorp F, Offringa R: **Strategies for immunotherapy of cancer.** *Adv Immunol* 2000, **75**:235-282.
- Grandics P: **Cancer: a single disease with a multitude of manifestations?** *J Carcinog* 2003, **2**:9.
- Anteunis A: **Cytochemical and ultrastructural studies concerning the cell coat glycoprotein in normal and transformed human blood lymphocytes. I. Variations of sialic acid containing glycoproteins subsequent to transformation of T and B lymphocytes by various kinds of stimulating agents.** *Exp Cell Res* 1974, **84**:31-39.
- Anteunis A, Vial M: **Cytochemical and ultrastructural studies concerning the cell coat glycoproteins in normal and transformed human blood lymphocytes. II. Comparison of lanthanum-retaining cell coat components in T and B lymphocytes transformed by various kinds of stimulating agents.** *Exp Cell Res* 1975, **90**:47-55.
- Bona C, Anteunis A: **Structure of the lymphocyte membrane. IV. Cell coat of lymphocytes obtained from various lymphoid organs in chicken and mice.** *Ann Immunol (Paris)* 1973, **124**:321-344.
- Jones BM: **A unifying hypothesis of cell adhesion.** *Nature* 1966, **212**:362-365.
- Doljanski F: **A new look at the cell surface.** *Isr J Med Sci* 1973, **9**:251-257.
- Santer V, Cone RE, Marchalonis JJ: **The glycoprotein surface coat on different classes of murine lymphocytes.** *Exp Cell Res* 1973, **79**:404-416.
- Bona C: **Physiological significance of the lymphocyte cell coat.** *Biomedicine* 1975, **22**:97-104.
- Gesner BM, Ginsburg V: **Effect of glycosidases on the fate of transfused lymphocytes.** *Proc Natl Acad Sci USA* 1964, **52**:750-755.
- Berney SN, Gesner BM: **The circulatory behaviour of normal and enzyme altered thymocytes in rats.** *Immunology* 1970, **18**:681-691.
- Woodruff J, Gesner BM: **The effect of neuraminidase on the fate of transfused lymphocytes.** *J Exp Med* 1969, **129**:551-567.
- Woodruff J, Gesner BM: **Lymphocytes: circulation altered by trypsin.** *Science* 1968, **161**:176-178.
- Lightbody JJ, Bach FH: **Cell mediated lympholysis: effect of papain on effector and target cells.** *Ann Immunol (Inst Pasteur)* 1973, **124**:311-319.
- Lindahl-Kiessling K, Peterson RD: **The mechanism of phytohemagglutinin (PHA) action II. The effect of certain enzymes and sugars.** *Exp Cell Res* 1969, **55**:81-84.
- Kemp RB: **Effect of the removal of cell surface sialic acids on cell aggregation « in vitro ».** *Nature* 1968, **218**:1255-1256.
- Douglas SD, Hoffman PF, Borjeson J, Chessin LN: **Studies on human peripheral blood lymphocytes in vitro. 3. Fine structural features of lymphocytes transformation by pokeweed mitogen.** *J Immunol* 1967, **98**:17-30.
- Bona C, Anteunis A, Robineaux R, Halpern B: **Structure of the lymphocyte membrane. 3. Chemical nature of the guinea pig lymphocyte membrane macromolecules reacting with heterologous ALS.** *Clin Exp Immunol* 1972, **12**:377-390.
- Currie GA, van Doorninck W, Bagshawe KD: **Effect of neuraminidase on the immunogenicity of early mouse tropoblast.** *Nature* 1968, **219**:191-192.

32. Ray PK, Gewurz H, Simmons RL: **The serologic behaviour of neuraminidase treated lymphoid cell. Alloantigenicity and complement sensitivity.** *Clin Exp Immunol* 1972, **11**:441-460.
33. Schlesinger M, Amos DB: **The effect of neuraminidase on the serological properties of murine lymphoid cells.** *Transpl Proc* 1971, **3**:895-897.
34. Simmons RL, Rios A, Ray PK: **Immunogenicity and antigenicity of lymphoid cells treated with neuraminidase.** *Nature New Biol* 1971, **231**:179-181.
35. Kaplan JG, Bona C: **Proteases as mitogens: the effect of trypsin and pronase on mouse and human lymphocytes.** *Exp Cell Res* 1974, **88**:388-394.
36. Flye MW, Grothaus EA, Amos DB: **Reactivity of human lymphoid cells following neuraminidase treatment.** *Surg Forum* 1971, **22**:97-99.
37. Gasic G, Berwick L: **Hale stain for sialic acid containing mucins. Adaptation to electron microscopy.** *J Cell Biol* 1963, **19**:223-228.
38. Rambourg A, Leblond CP: **Electronmicroscope observation on carbohydrate-rich cell coat present at the surface of cells in the rat.** *J Cell Biol* 1967, **32**:27-53.
39. Lichtman MA, Weed RI: **Electrophoretic mobility and N-acetyl neuraminic acid content of human normal and leukemic lymphocytes and granulocytes.** *Blood* 1970, **35**:12-22.
40. Abercrombie M, Ambrose EJ: **The surface properties of cancer cells: a review.** *Cancer Res* 1962, **22**:525-548.
41. Calman F: **Ultrastructural comparison of the cell coat in normal and chronic lymphocytic leukaemic blood lymphocytes by Concanavalin A labelling and cationic staining.** *Pathol Eur* 1975, **10**:203-214.
42. Mallucci L, Poste GH, Wells V: **Synthesis of cell coat in normal and transformed cells.** *Nat New Biol* 1972, **235**:222-223.
43. Gasic G, Loebel F: **Cytochemical identification of protein amino acids in the cell coat of mouse ascites tumor cells.** *Lab Invest* 1966:1310-1319.
44. Rittenhouse HG, Rittenhouse JW, Takemoto L: **Characterization of the cell coat of Ehrlich ascites tumor cells.** *Biochemistry* 1978, **17**:829-837.
45. Kovacs A: **Process for the preparations of anti-tumor therapeutics.** *Hungarian Patent No. 200093 B* 1983.
46. Gasic G, Gasic T: **Removal of sialic acid from the cell coat of tumor cells and vascular endothelium, and its effect on metastasis.** *Proc Natl Acad Sci USA* 1962, **48**:1172-1177.
47. Bagshawe KD, Currie GA: **Immunogenicity of L 1210 murine leukemia cells after treatment with neuraminidase.** *Nature* 1968, **218**:1254-1255.
48. Sanford BH: **An alteration in tumor histocompatibility induced by neuraminidase.** *Transplantation* 1967, **5**:1273-1279.
49. Gasic G, Gasic T: **Removal of PAS positive surface sugars in tumor cells by glycosidases.** *Proc Soc Exp Biol Med* 1963, **114**:660-663.
50. Gasic G, Gasic T: **Removal and regeneration of the cell coating of tumour cells.** *Nature* 1962, **196**:170.
51. Anghileri LJ, Dermietzel R: **Cell coat in tumor cells-Effects of trypsin and EDTA: A biochemical and morphological study.** *Oncology* 1976, **33**:17-23.
52. Gately MK, Glaser M, McCarron RM, Dick SJ, Dick MD, Mettetal RW, Kornblith PL: **Mechanisms by which human gliomas may escape cellular immune attack.** *Acta Neurochir* 1982, **64**:175-197.
53. Dick SJ, Macchi B, Papazoglou S, Oldfield EH, Kornblith PL, Smith BH, Gately MK: **Lymphoid cell-glioma cell interaction enhances cell coat production by human gliomas: novel suppressor mechanism.** *Science* 1983, **220**:739-742.
54. McBride WH, Bard JB: **Hyaluronidase sensitive halos around adherent cells. Their role in blocking lymphocyte-mediated cytotoxicity.** *J Exp Med* 1979, **149**:507-515.
55. Thorling EB, Larsen B, Nielsen H: **Inhibitory effect of DEAE-dextran on tumour growth. 3. Effect of charge density and molecular size.** *Acta Path Microbiol Scand A* 1971, **79**:81-90.
56. Marquez M, Nilsson S, Lennartsson L, Liu Z, Tammela T, Raitanen M, Holmberg AR: **Charge-dependent targeting: Results in six tumor cell lines.** *Anticancer Res* 2004, **24**:1347-1352.
57. Opal SM: **Interactions between coagulation and inflammation.** *Scand J Infect Dis* 2003, **35**:545-554.
58. Semeraro N, Lattanzio A, Montemurro P, Papanice M, De Lucia O, De Bellis G, Giordano D: **Mechanisms of blood clotting activation in inflammation: the role of mononuclear phagocytes.** *Int J Tissue React* 1985, **7**:313-320.
59. Helin H: **Macrophage procoagulant factors – mediators of inflammatory and neoplastic tissue lesions.** *Med Biol* 1986, **64**:167-176.
60. Rauch U, Bonderman D, Bohrmann B, Badimon JJ, Himber J, Riederer MA, Nemerson Y: **Transfer of tissue factor from leukocytes to platelets is mediated by CD15 and tissue factor.** *Blood* 2000, **96**:170-175.
61. Bevers EM, Comfurius P, Dekkers DW, Harmsma M, Zwaal RF: **Transmembrane phospholipid distribution in blood cells: control mechanisms and pathophysiological significance.** *Biol Chem* 1989, **379**:973-986.
62. Okajima K: **Regulation of inflammatory responses by natural anticoagulants.** *Immunol Rev* 2001, **184**:258-274.
63. Oelschläger C, Romisch J, Staubitz A, Stauss H, Leithäuser B, Tillmanns H, Holschermann H: **Antithrombin III inhibits nuclear factor κ B activation in human monocytes and vascular endothelial cells.** *Blood* 2002, **99**:4015-4020.
64. Sturn DH, Kaneider NC, Feistritz C, Djanani A, Fukudome K, Wiederemann CJ: **Expression and function of the endothelial protein C receptor in human neutrophils.** *Blood* 2003, **102**:1499-1505.
65. Isobe H, Okajima K, Uchiba M, Mizutani A, Harada N, Nagasaki A, Okabe K: **Activated protein C prevents endotoxin-induced hypotension in rats by inhibiting excessive production of nitric oxide.** *Circulation* 2001, **104**:1171-1175.
66. Joyce DE, Gelbert L, Ciaccia A, DeHoff B, Grinnell BW: **Gene expression profile of antithrombotic protein C defines new mechanisms modulating inflammation and apoptosis.** *J Biol Chem* 2001, **276**:11199-11203.
67. Burstein SA: **Cytokines, platelet production and hemostasis.** *Platelets* 1997, **8**:93-104.
68. Pendurthi UR, Alok D, Rao LV: **Binding of factor VIIa to tissue factor induces alterations in gene expression in human fibroblast cells: upregulation of poly(A) polymerase.** *Proc Natl Acad Sci USA* 1997, **94**:12598-12603.
69. Miller DL, Yaron R, Yellin MJ: **CD40L-CD40 interactions regulate endothelial cell surface tissue factor and thrombomodulin expression.** *J Leukoc Biol* 1998, **63**:373-379.
70. Andre P, Prasad KS, Denis CV, He M, Papalia JM, Hynes RO, Phillips DR, Wagner DD: **CD40L stabilizes arterial thrombi by a b3 integrin-dependent mechanism.** *Nat Med* 2002, **8**:247-252.
71. Henn V, Slupsky JR, Grafe M, Anagnostopoulos I, Forster R, Muller-Berghaus G, Kroccek RA: **CD40 ligand on activated platelets triggers an inflammatory reaction of endothelial cells.** *Nature* 1998, **391**:591-594.
72. Trousseau A: **Phlegmasia alba dolens.** In *Clinique Medicale de l'Hotel-Dieu de Paris Volume 3*. 2nd edition. Balliere, Paris; 1865:654-712.
73. Rickles FR, Edwards RL: **Activation of blood coagulation in cancer: Trousseau's syndrome revisited.** *Blood* 1983, **62**:14-31.
74. Donati MB: **Cancer and thrombosis: from phlegmasia alba dolens to transgenic mice.** *Thromb Haemost* 1995, **74**:278-281.
75. Gordon SG: **Cancer cell procoagulants and their role in malignant disease.** *Semin Thromb Haemost* 1992, **18**:424-433.
76. Rao LV: **Tissue factor as a tumor procoagulant.** *Cancer Metastasis Rev* 1992, **11**:249-266.
77. Andoh D, Kubota T, Takada M, Tanaka H, Kobayashi N, Maekawa T: **Tissue factor activity in leukaemia cells. Special reference to disseminated intravascular coagulation.** *Cancer* 1987, **59**:748-54.
78. Nemerson Y: **The tissue factor pathway of blood coagulation.** *Semin Haematol* 1992, **29**:170-176.
79. Gordon SG, Mourad AM: **The site of activation of factor X by cancer procoagulant.** *Blood Coagul Fibrinolysis* 1991, **2**:735-739.
80. Falanga A, Gordon SG: **Isolation and characterization of cancer procoagulant: a cysteine protease from malignant tissue.** *Biochemistry* 1985, **24**:5558-5567.
81. Donati MB, Falanga A, Consonni R, Alessio MG, Bassan R, Buelli M, Borin L, Catani L, Pogliani E, Gugliotti L: **Cancer procoagulant in acute non lymphoid leukaemia: relationship of enzyme detection to disease activity.** *Thromb Haemost* 1990, **64**:11-16.
82. Kwaan HC, Keer HN: **Fibrinolysis and cancer.** *Semin Thromb Haemost* 1990, **16**:230-235.

83. Falanga A, Marchetti M, Giovanelli S, Barbui T: **All-trans-retinoic acid counteracts endothelial cell procoagulant activity induced by a human promyelocytic leukaemia-derived cell line (NB4).** *Blood* 1996, **87**:613-617.
84. Gianni M, Norio P, Terao M, Falanga A, Marchetti M, Rambaldi A, Garrattini E: **The effect of dexamethasone on proinflammatory cytokine expression, cell growth and maturation during granulocytic differentiation of acute promyelocytic leukaemic cells.** *Eur Cytokine Netw* 1995, **6**:157-165.
85. Honn KV, Tang DG, Chen YQ: **Platelets and cancer metastasis: more than an epiphenomenon.** *Semin Thromb Haemost* 1992, **18**:392-415.
86. Marchetti M, Falanga A, Giovanelli S, Oldani E, Barbui T: **All-trans-retinoic acid increases the adhesion to endothelium of the acute promyelocytic leukaemia cell line NB4.** *Br J Haematol* 1996, **93**:360-366.
87. Rickles FR, Edwards RL: **Leukocytes and tumour cells in thrombosis.** In *Haemostasis and Thrombosis: Basic Principles and Clinical Practice* Edited by: Colman RW, Hirsh J, Marder VJ, Salzman EV. Lippincott, Philadelphia, PA, USA; 1994:1164-1179.
88. O'Meara RAQ: **Coagulative properties of cancer.** *Irish J Med Sci* 1958, **6**:474-479.
89. Dvorak HF, Senger DR, Dvorak AM: **Fibrin as a component of the tumor stroma: origins and biological significance.** *Cancer Metastasis Rev* 1983, **2**:41-73.
90. Hiramoto R, Bernecky J, Jurandowski J: **Fibrin in human tumors.** *Cancer Res* 1960, **20**:592-593.
91. Dvorak HF: **Tumor: wounds that do not heal.** *N Engl J Med* 1986, **315**:1650-1659.
92. Colvin RB, Dvorak HF: **Fibrinogen/fibrin on the surface of macrophages: detection, distribution binding requirements, and possible role in the macrophage adherence phenomena.** *J Exp Med* 1975, **142**:1377-1390.
93. Gunji Y, Gorelik E: **Role of fibrin coagulation in protection of murine tumor cells from destruction by cytotoxic cells.** *Cancer Res* 1988, **48**:5216-5221.
94. Gorelik E: **Augmentation of the antimetastatic effect of anticoagulant drugs by immunostimulation in mice.** *Cancer Res* 1987, **47**:809-815.
95. Dvorak HF, Dvorak AM, Manseau EJ, Wiberg L, Churchill WH: **Fibrin gel investment associated with line I and line 10 solid tumor growth, angiogenesis, and fibroplasia in guinea pigs. Role of cellular immunity, myofibroblasts, microvascular damage, and infarction in line I tumor regression.** *J Natl Cancer Inst* 1979, **62**:1459-1472.
96. Dittman WA, Majerus PV: **Structure and function of thrombomodulin: a natural anticoagulant.** *Blood* 1990, **75**:329-336.
97. Moore KL, Esmon CT, Esmon NL: **Tumor necrosis factor leads to the internalization and degradation of thrombomodulin from the surface of bovine aortic endothelial cells in culture.** *Blood* 1989, **73**:159-165.
98. Qi J, Kreutzer DL: **Fibrin activation of vascular endothelial cells: induction of IL - 8 expression.** *J Immunol* 1995, **155**:867-876.
99. Contrino J, Hair G, Kreutzer DL, Rickles FR: **In situ detection of tissue factor in vascular endothelial cells: correlation with the malignant phenotype of breast disease.** *Nat Med* 1996, **2**:209-215.
100. Koomagi R, Volm M: **Tissue factor expression in human non-small-cell lung carcinoma measured by immunohistochemistry: correlation between tissue factor and angiogenesis.** *Int J Cancer* 1998, **79**:19-22.
101. Abdulkadir SA, Carvalhal GF, Kaleem Z, Kiesel W, Humphrey PA, Catalona WJ, Milbrandt J: **Tissue Factor expression and angiogenesis in human prostate carcinoma.** *Hum Pathol* 2000, **31**:443-447.
102. Honn KV, Tang DG, Chen YQ: **Platelets and cancer metastasis: more than an epiphenomenon.** *Semin Thromb Haemost* 1992, **18**:392-415.
103. Schwartz GK: **Invasion and metastases in gastric cancer: in vitro and in vivo models with clinical correlations.** *Semin Oncol* 1996, **23**:316-324.
104. Orr FW, Wang HH: **Tumor cell interactions with the microvasculature: a rate-limiting step in metastasis.** *Surg Oncol Clin N Am* 2001, **10**:357-381.
105. Hara Y, Steiner M, Baldini MG: **Characterization of the platelet-aggregating activity of tumour cells.** *Cancer Res* 1980, **40**:1217-1222.
106. Nathan CF: **Mechanism of macrophage antimicrobial activity.** *Trans R Soc Trop Med Hyg* 1983, **77**:620-630.
107. Unanue ER: **Cooperation between mononuclear phagocytes and lymphocytes in immunity.** *New Engl J Med* 1980, **303**:977-985.
108. van Furth R: **Mononuclear phagocytes. Characteristics, Physiology, and Function.** Martinus Nijhoff Publishers; 1985.
109. Semararo N, Colucci M: **Tissue factor in health and disease.** *Thromb Haemost* 1997, **78**:759-764.
110. Lorenzet R, Peri G, Locati D, Allavena P, Colucci M, Semararo N, Mantovani A, Donati MB: **Generation of procoagulant activity by mononuclear phagocytes: a possible mechanism contributing to blood clotting activation within malignant tissue.** *Blood* 1983, **62**:271-273.
111. Mussoni L, Donati MB: **Expression of plasminogen activator as a marker of stimulation in tumour-associated macrophages.** *Haemostasis* 1988, **18**:66-71.
112. Gastpar H: **Platelet-cancer cell interaction in metastasis formation: a possible therapeutic approach to metastasis prophylaxis.** *J Med* 1977, **8**:103-114.
113. Karparkin S, Pearlstein E: **Heterogeneous mechanisms of tumor cell-induced platelet aggregation with possible pharmacological strategy toward prevention of metastases.** In *Hemostatic Mechanisms and Metastases* Edited by: Honn KV, Sloane BF. Martinus Nijhoff, Boston; 1984:139-169.
114. Gasic GJ, Gasic TB, Stewart GJ: **Mechanisms of platelet aggregation by murine tumor cell shedding.** In *Hemostatic Mechanisms and Metastases* Edited by: Honn KV, Sloane BF. Martinus Nijhoff, Boston; 1984:127-138.
115. Falanga A, Rickles FR: **Pathophysiology of the thrombophilic state in the cancer patient.** *Semin Thromb Haemost* 1999, **25**:173-182.
116. Gasic GJ, Gasic TB, Stewart CC: **Antimetastatic effects associated with platelet reduction.** *Proc Natl Acad Sci USA* 1968, **61**:46-52.
117. Gasic GJ, Gasic TB, Galanti N, Johnson T, Murphy S: **Platelet-tumor cell interaction in mice. The role of platelets in the spread of malignant disease.** *Int J Cancer* 1973, **11**:704-718.
118. Honn KV, Cicone B, Skoff A: **Prostacyclin: a potent antimetastatic agent.** *Science* 1981, **212**:1270-1272.
119. Honn KV, Cavanaugh P, Evens C, Taylor JD, Sloane BF: **Tumor cell-platelet aggregation: induced by cathepsin B-like proteinase and inhibited by prostacyclin.** *Science* 1982, **217**:540-542.
120. Gasic GJ, Gasic TB, Murphy S: **Anti-metastatic effect of aspirin.** *Lancet* 1972, **2**:932-933.
121. Thun MJ, Namboodiri MM, Heath CW Jr: **Aspirin use and reduced risk of fatal colon cancer.** *N Engl J Med* 1991, **325**:1593-1596.
122. Fuchs C: **Aspirin, COX-2 Inhibitors Effective as Adjuvant Therapy in Stage III Colon Cancer.** *ASCO 2005 Annual Meeting: Abstract 3530*.
123. Kune GA, Kune S, Watson LF: **Colorectal cancer risk, chronic illnesses, operations, and medications: case control results from the Melbourne Colorectal Cancer Study.** *Cancer* 1988, **48**:4399-4404.
124. Labayle D, Fischer D, Vielh P, Drouhin F, Pariente A, Borjes C, Duhamel O, Troussset M, Attali P: **Sulindac causes regression of rectal polyps in familial adenomatous polyposis.** *Gastroenterology* 1991, **101**:635-639.
125. Pollard M, Luckert PH: **Treatment of chemically induced intestinal cancers with indomethacin.** *Proc Soc Exp Biol Med* 1981, **167**:161-164.
126. Clifton EE, Grossi CE: **The effect of human plasmin on the toxic properties and growth of the VX2 carcinoma and the Brown Pearce carcinoma of rabbits.** *Cancer* 1956, **9**:1147-1152.
127. Clifton EE: **Fibrinolytic therapy for thrombo-embolic disease: principles and practice.** *J La State Med Soc* 1966, **118**:309-319.
128. Clifton EE: **Effect of fibrinolysin on spread of cancer.** *Fed Proc* 1966, **25**:89-93.
129. Rudenstan CM: **Effect of fibrinolytic antifibrinolytic and flow promoting agents in metastases formation and tumor growth.** In *Endogenous factors influencing host tumor balance* University of Chicago Press, Chicago (IL); 1967:277-298.

130. Clifton EE, Agostino D: **Effect of inhibitors of fibrinolytic enzymes on development of pulmonary metastases.** *J Natl Cancer Inst* 1964, **33**:753-763.
131. Luzzatto G, Schafer AI: **The prethrombotic state in cancer.** *Semin Oncol* 1990, **17**:147-158.
132. Lee AY, Levine MN: **The thrombophilic state induced by therapeutic agents in cancer patients.** *Semin Thromb Haemost* 1999, **25**:137-145.
133. Kakkar VV, Howe CT, Nicolaidis AN, Renney JT, Clarke MB: **Deep vein thrombosis of the leg. Is there a "high risk" group?** *Am J Surg* 1970, **120**:527-530.
134. Clagett GP, Anderson FA, Geerts W, Heit JA, Knudson M, Lieberman JR, Merli GJ, Wheeler HB: **Prevention of venous thromboembolism.** *Chest* 1998, **114**(Suppl):531S-560S.
135. Pritchard KI, Paterson AH, Paul NA, Zee B, Fine S, Pater J: **Increased thromboembolic complications with concurrent tamoxifen and chemotherapy in a randomized trial of adjuvant therapy for women with breast cancer. National Cancer Institute of Canada Clinical Trials Group Breast Cancer Site Group.** *J Clin Oncol* 1996, **14**:2731-2737.
136. Saphner T, Tormey DC, Gray R: **Venous and arterial thrombosis in patients who received adjuvant chemotherapy for breast cancer.** *J Clin Oncol* 1991, **9**:286-294.
137. Goodnough LT, Saito H, Manni A, Jones PK, Pearson OH: **Increased incidence of thrombosis in stage IV breast cancer patients treated with a five-drug chemotherapy regimen A study of 159 patients.** *Cancer* 1984, **54**:1264-1268.
138. Wall JG, Weiss RB, Norton L, Perloff M, Rice MA, Korzun AH, Wood WC: **Arterial thrombosis associated with adjuvant chemotherapy for breast cancer: a Cancer and Leukaemia Group B study.** *Am J Med* 1989, **87**:501-504.
139. Falanga A: **Mechanisms of hypercoagulation in malignancy and during chemotherapy.** *Haemostasis* 1998, **28**(Suppl S3):50-60.
140. Levine M, Hirsh J, Gent M, Arnold A, Warr D, Falanga A, Samosh M, Bramwell V, Pritchard KI, Stewart D, et al.: **Double-blind randomized trial of very-low-dose warfarin for prevention of thromboembolism in stage IV breast cancer.** *Lancet* 1994, **343**:886-889.
141. Barbui T, Finazzi G, Grassi A, Marchioli R: **Thrombosis in cancer patients treated with hematopoietic growth factors - a meta-analysis. On behalf of the Subcommittee on Haemostasis and Malignancy of the Scientific and Standardization Committee of the ISTH.** *Thromb Haemost* 1996, **75**:368-371.
142. Falanga A, Marchetti M, Evangelista V, Manarini S, Oldani E, Giovanelli S, Galbusera M, Cerletti C, Barbui T: **Neutrophil activation and hemostatic changes in healthy donors given granulocyte-colony stimulating factor.** *Blood* 1999, **93**:2506-2514.
143. Sorensen HT, Mellekjær L, Steffensen FH, Olsen JH, Nielsen GL: **The risk of a diagnosis of cancer after primary deep venous thrombosis or pulmonary embolism.** *N Engl J Med* 1998, **338**:1169-1173.
144. Baron JA, Gridley G, Weiderpass E, Nyren O, Linet M: **Venous thromboembolism and cancer.** *Lancet* 1998, **351**:1077-1080.
145. Kwapinski G, Oliver H, Kwapinski E, Stein M: **Microbial-like antigens in human leukemia.** *Oncology* 1978, **35**:263-266.
146. Rocken C, Carl-McGrath S: **Pathology and pathogenesis of hepatocellular carcinoma.** *Dig Dis* 2001, **19**:269-278.
147. Dammacco F, Sansonno D, Piccoli C, Racanelli V, D'Amore FP, Lauletta G: **The lymphoid system in hepatitis C virus infection: autoimmunity, mixed cryoglobulinemia, and Overt B-cell malignancy.** *Semin Liver Dis* 2000, **20**:143-157.
148. Vardareli E, Saricam T, Isiksoy S, Yavuz H, Ozakyol A, Kircali B: **Type I gastric carcinoid tumor: another extrahepatic manifestation of hepatitis C virus infection?** *Turk J Gastroenterol* 2003, **14**:194-196.
149. Ramos-Casals M, Trejo O, Garcia-Carrasco M, Cervera R, De La Red G, Gil V, Lopez-Guillermo A, Ingelmo M, Font J: **Triple association between hepatitis C virus infection, systemic autoimmune diseases, and B cell lymphoma.** *J Rheumatol* 2004, **31**:495-499.
150. Hermine O, Lefrere F, Bronowicki JP, Mariette X, Jondeau K, Eclache-Saudreau V, Delmas B, Valensi F, Cacoub P, Brechot C, Varet B, Troussard X: **Regression of splenic lymphoma with villous lymphocytes after treatment of hepatitis C virus infection.** *N Engl J Med* 2002, **347**:89-94.
151. Dorak MT: **The implications for childhood leukemia of infection with adenovirus.** *Trends Microbiol* 1996, **4**:60-63.
152. Mueller N, Hinkula J, Wahren B: **Elevated antibody titers against cytomegalovirus among patients with testicular cancer.** *Int J Cancer* 1988, **41**:399-403.
153. D'Elia MM, Amedei A, Benagiano M, Azzurri A, Del Prete G: **Helicobacter pylori, T cells and cytokines: the "dangerous liaisons".** *FEMS Immunol Med Microbiol* 2005, **44**:113-119.
154. Gupta V, Eden AJ, Mills MJ: **Helicobacter pylori and autoimmune neutropenia.** *Clin Lab Haematol* 2002, **24**:183-185.
155. Vilchez RA, Butel JS: **SV40 in human brain cancers and non-Hodgkin's lymphoma.** *Oncogene* 2003, **22**(33):5164-5172.
156. Ferreri AJ, Ponzoni M, Guidoboni M, De Conciliis C, Resti AG, Mazzi B, Lettini AA, Demeter J, Dell'Oro S, Doglioni C, Villa E, Boiocchi M, Dolcetti R: **Regression of ocular adnexal lymphoma after Chlamydia psittaci-eradicating antibiotic therapy.** *J Clin Oncol* 2005, **23**(22):5067-5073.
157. Kozliuk VA, Lakatos VP, Kozliuk AS, Proshchakov KV, Kozar MI: **Cervicitis and cervical intraepithelial neoplasia: cytomorphological and molecular biology analysis.** *Lik Sprava* 2003, **8**:29-36.
158. Guijon FB, Paraskevas M, Brunham R: **The association of sexually transmitted diseases with cervical intraepithelial neoplasia: a case-control study.** *Am J Obstet Gynecol* 1985, **151**:185-190.
159. Bezuglova TV, Aspiz NV: **Role of Mycoplasma in the processes of abnormal growth.** *Vestn Akad Med Nauk SSSR* 1986, **1**:76-82.
160. Averna R, Martelli D, Migliorini D, Saudelli M: **Mycoplasmas and dysplasia of the uterine cervix.** *Boll Ist Sieroter Milan* 1980, **59**:348-358.
161. Cassell GH, Cole BC: **Mycoplasmas as agents of human disease.** *N Engl J Med* 1981, **304**:80-89.
162. Taylor Robinson D, McCormack WM: **The genital mycoplasmas.** *N Engl J Med* 1980, **302**:1003-1010.
163. Cole BC, Cassell GH: **Mycoplasma infections as models of chronic joint inflammation.** *Arthritis Rheum* 1979, **22**:1375-1381.
164. Barile MF, Yoshida H, Roth H: **Rheumatoid arthritis: new findings on the failure to isolate or detect mycoplasmas by multiple cultivation or serologic procedures and a review of the literature.** *Rev Infect Dis* 1991, **13**:571-582.
165. Fiberg J: **Mycoplasmas and ureaplasmas in reproductive failure.** *Contemp Obstet Gynecol* 1983, **28**:271-286.
166. Rakovskaia IV, Gorina LG: **Co-leukemogenic activity of protein preparations, isolated from cells of Mycoplasma arthritidis.** *Vestn Akad Med Nauk SSSR* 1985, **10**:62-66.
167. Rhew DC, Gaultier CR, Daar ES, Zakowski PC, Said J: **Infections in patients with chronic adult T-cell leukemia/lymphoma: case report and review.** *Clin Infect Dis* 1995, **21**:1014-1016.
168. Krepler P: **Infections in children with malignant disease.** *Wien Klin Wochenschr* 1979, **91**:707-715.
169. Schmidhauser C, Dudler R, Schmidt T, Parish RW: **A mycoplasma protein influences tumour cell invasiveness and contact inhibition in vitro.** *J Cell Sci* 1990, **95**:499-506.
170. Chan PJ, Seraj IM, Kalugdan TH, King A: **Prevalence of mycoplasma conserved DNA in malignant ovarian cancer detected using sensitive PCR-ELISA.** *Gynecol Oncology* 1996, **63**:258-260.
171. Sasaki H, Igaki H, Ishizuka T, Kogoma Y, Sugimura T, Terada M: **Presence of streptococcus DNA sequence in surgical specimens of gastric cancer.** *Jpn J Cancer Res* 1995, **86**:791-794.
172. Huang S, Li SJ, Wu J, Meng L, Shou CC: **Mycoplasma infections and different human carcinomas.** *World J Gastroenterol* 2001, **7**:266-269.
173. Paton GR, Jacobs JP, Perkins FT: **Chromosome changes in human Diploid-cell cultures infected with mycoplasma.** *Nature* 1965, **207**:43-45.
174. Bezuglova TV, Lange A, Gusman BS, Ritter E: **Development of lung tumors in Syrian hamsters with a mixed Mycoplasma pneumoniae and influenza virus infection.** *Biull Eksp Biol Med* 1985, **99**:476-477.
175. Tsai S, Wear DJ, Shih JW, Lo SC: **Mycoplasmas and oncogenesis: Persistent infection and multistage malignant transformation.** *Proc Natl Acad Sci USA* 1995, **92**:10197-10201.
176. Rawadi G, Roman-Roman S, Castedo M, Dutilleul V, Susin S, Marchetti P, Geuskens M, Kroemer G: **Effects of Mycoplasma fermentans on the myelomonocytic lineage: Different molecular entities with cytokine inducing and cytotoxic potential.** *J Immunol* 1996, **156**:670-678.

177. Mihai G, Netea J, van der Meer WM, Kullberg B-J: **Toll-like receptors as an escape mechanism from the host defense.** *Trends in Microbiol* 2004, **12**:484-488.
178. Kopp E, Medzhitov R: **Recognition of microbial infection by Toll-like receptors.** *Curr Opin Immunol* 2003, **15**:396-401.
179. Akira S, Hemmi H: **Recognition of pathogen-associated molecular patterns by TLR family.** *Immunol Lett* 2003, **85**:85-95.
180. Takeuchi O, et al.: **Discrimination of bacterial lipoproteins by Toll-like receptor 6.** *Int Immunol* 2001, **13**:933-940.
181. Choy EH, Panayi GS: **Cytokine pathways and joint inflammation in rheumatoid arthritis.** *N Engl J Med* 2001, **344**:907-916.
182. Lucas K, Hohlfeld R: **Differential aspects of cytokines in the immunopathology of multiple sclerosis.** *Neurology* 1995, **45**:S4-S5.
183. Arnett HA, Mason J, Marino M, Suzuki K, Matsushima GK, Ting JP: **TNFalpha promotes proliferation of oligodendrocyte progenitors and remyelination.** *Nat Neurosci* 2001, **4**:1116-1122.
184. Linker-Israeli M: **Cytokine abnormalities in human lupus.** *Clin Immunol Immunopathol* 1992, **63**:10-12.
185. Moqattash S, Lutton JD: **Leukemia cells and the cytokine network.** *Proc Soc Exp Biol Med* 1998, **219**:8-27.
186. Lattime EC, Mastrangelo MJ, Bagasra O, Li W, Berd D: **Expression of cytokine mRNA in human melanoma tissues.** *Cancer Immunol Immunother* 1995, **41**:151-156.
187. Kruger-Krasagakes S, Krasagakis K, Garbe C, Schmitt E, Huls C, Blankenstein T, Diamantstein T: **Expression of interleukin-10 in human melanoma.** *Br J Cancer* 1994, **70**:1182-1185.
188. Huang M, Wang J, Lee P, Sharma S, Mao JT, Meissner H, Uyemura K, Modlin R, Wollman J, Dubinett SM: **Human non-small cell lung cancer cells express a type 2 cytokine pattern.** *Cancer Res* 1995, **55**:3847-3853.
189. Nakagomi H, Pisa P, Pisa EK, Yamamoto Y, Halapi E, Backlin K, Juhlin C, Kiessling R: **Lack of interleukin-2 (IL-2) expression and selective expression of IL-10 mRNA in human renal cell carcinoma.** *Int J Cancer* 1995, **63**:366-371.
190. Lattime EC, McCue PA, Keely FX, Li W, Gomella LG: **Expression of IL-10 mRNA in biopsies of superficial and invasive TCC of the human bladder.** *Proc Am Assoc Cancer Res* 1995, **36**:462.
191. Sato T, McCue P, Masuoka K, Salwen S, Lattime EC, Mastrangelo MJ, Berd D: **Interleukin 10 production by human melanoma.** *Clin Cancer Res* 1996, **2**:1383-1390.
192. Rico MJ, Matar P, Gervasoni SI, Bonfil RD, Calcaterra N, Scharovsky OG: **The transition to the metastatic phenotype of rat lymphoma cells involves up-regulation of IL-10 receptor expression and IL-10 secretion.** *Clin Exp Metastasis* 2005, **22**:127-135.
193. Pellegrino A, Vacca A, Scavelli C, Dammacco F: **Chemokines and tumors.** *Recenti Prog Med* 2002, **93**:642-654.
194. Fassone L, Gaidano G, Ariatti C, Vivenza D, Capello D, Gloghini A, Cilia AM, Buonaiuto D, Rossi D, Pastore C, Carbone A, Saglio G: **The role of cytokines in the pathogenesis and management of AIDS-related lymphomas.** *Leuk Lymphoma* 2000, **38**:481-488.
195. Kurebayashi J: **Regulation of interleukin-6 secretion from breast cancer cells and its clinical implications.** *Breast Cancer* 2000, **7**:124-129.
196. Bar-Eli M: **Role of interleukin-8 in tumor growth and metastasis of human melanoma.** *Pathobiology* 1999, **67**:12-18.
197. Sparmann A, Bar-Sagi D: **Ras-induced interleukin-8 expression plays a critical role in tumor growth and angiogenesis.** *Cancer Cell* 2004, **6**:447-458.
198. Skinnider BF, Kapp U, Mak TW: **Interleukin 13: a growth factor in Hodgkin lymphoma.** *Int Arch Allergy Immunol* 2001, **126**:267-276.
199. Kirkbride KC, Blobe GC: **Inhibiting the TGF-b signaling pathway as a means of cancer immunotherapy.** *Expert Opin Biol Ther* 2003, **3**:251-261.
200. Lai R, O'Brien S, Maushouri T, Rogers A, Kantarjian H, Keating M, Albitar M: **Prognostic value of plasma interleukin-6 levels in patients with chronic lymphocytic leukemia.** *Cancer* 2002, **95**:1071-1075.
201. Fortis C, Foppoli M, Gianotti L, Galli L, Citterio G, Consogno G, Gentilini O, Braga M: **Increased interleukin-10 serum levels in patients with solid tumors.** *Cancer Lett* 1996, **104**:1-5.
202. Nabioullin R, Sone S, Mizuno K, Yano S, Nishioka Y, Haku T, Ogura T: **Interleukin-10 is a potent inhibitor of tumor cytotoxicity by human monocytes and alveolar macrophages.** *J Leukocyte Biol* 1994, **55**:437-442.
203. Ding L, Linsley PS, Huang LY, Germain RN, Shevach EM: **IL-10 inhibits macrophage costimulatory activity by selectively inhibiting the up-regulation of B7 expression.** *J Immunol* 1993, **151**:1224-1234.
204. de Waal Malefyt R, Haanen J, Spits H, Roccarolo MG, te Velde A, Figdor C, Johnson K, Kastelstein R, Yssel H, de Vries JE: **Interleukin-10 (IL-10) and viral IL-10 strongly reduce antigen-specific human T cell proliferation by diminishing the antigen-presenting capacity of monocytes via downregulation of class II major histocompatibility complex expression.** *J Exp Med* 1991, **174**:915-924.
205. Willems F, Marchant A, Delville JP, Gerard C, Delvaux A, Velu T, de Boer M, Goldman M: **Interleukin-10 inhibits B7 and intercellular adhesion molecule-1 expression on human monocytes.** *Eur J Immunol* 1994, **24**:1007-1009.
206. Rohrer JW, Coggin JH: **CD81 T cell clones inhibit antitumor T cell function by secreting IL-10.** *J Immunol* 1995, **155**:5719-5727.
207. Afford S, Randhawa S: **Apoptosis.** *Mol Path* 2002, **53**:55-63.
208. Yuan XM, Li W, Dalen H, Lotem J, Kama R, Sachs L, Brunk UT: **Lysosomal destabilization in p53-induced apoptosis.** *Proc Natl Acad Sci USA* 2002, **99**:6286-6291.
209. Pellegris G, Ravagnani F, Notti P, Fissi S, Lombardo C: **B and C hepatitis viruses, HLA-DQ1 and -DR3 alleles and autoimmunity in patients with hepatocellular carcinoma.** *J Hepatol* 2002, **36**:521-526.
210. Bolognini G, Riva G: **Lymphoproliferative diseases and paraproteinemias in Sjogren's syndrome.** *Schweiz Med Wochenschr* 1975, **105**:1493-1505.
211. Agnello V: **The aetiology of mixed cryoglobulinaemia associated with hepatitis C virus infection.** *Scand J Immunol* 1995, **42**:179-184.
212. Dammacco F, Sansonno D, Cornacchiulo V, Mennuni C, Carbone R, Lauletta G, Iacobelli AR, Rizzi R: **Hepatitis C virus infection and mixed cryoglobulinemia: A striking association.** *Int J Clin Lab Res* 1993, **23**:45-49.
213. Andreev VC, Zlatkov NB: **Systemic lupus erythematosus and neoplasia of the lymphoreticular system.** *Brit J Derm* 1968, **80**:503-508.
214. Lewis RB, Castor CW, Kinsley RE, Bole GG: **Frequency of neoplasia in systemic lupus erythematosus and rheumatoid arthritis.** *Arthritis Rheum* 1976, **19**:1256-1260.
215. Till M, Rapson N, Smith PG: **Family studies in acute leukaemia in childhood: a possible association with autoimmune disease.** *Br J Cancer* 1979, **40**:62-71.
216. Venables P: **Epstein-Barr virus infection and autoimmunity in rheumatoid arthritis.** *Ann Rheum Dis* 1988, **47**:265-269.
217. Ambinder R: **Infection and lymphoma.** *N Engl J Med* 2003, **349**:1309-1311.
218. Hiemstra HS, Schloot NC, van Veelen PA, Willemen SJ, Franken KL, van Rood JJ, de Vries RR, Chaudhuri A, Behan PO, Drijfhout JW, Roep BO: **Cytomegalovirus in autoimmunity: T cell crossreactivity to viral antigen and autoantigen glutamic acid decarboxylase.** *Proc Natl Acad Sci USA* 2001, **98**:3988-3991.
219. Cammarata RJ, Rodnan GP, Jensen VVN: **Systemic rheumatic disease and malignant lymphoma.** *Arch Intern Med* 1963, **111**:330-337.
220. Groux H, Cottrez F: **The complex role of interleukin-10 in autoimmunity.** *J Autoimmun* 2003, **20**:281-285.
221. Burnham TK: **Antinuclear antibodies in patients with malignancies.** *Lancet* 1972, **2(7774)**:436.
222. Abu-Shakra M, Buskila D, Ehrenfeld M, Conrad K, Shoenfeld Y: **Cancer and autoimmunity: autoimmune and rheumatic features in patients with malignancies.** *Ann Rheum Dis* 2001, **60**:433-441.
223. Mills JA: **Connective tissue disease associated with malignant neoplastic disease.** *J Chronic Dis* 1963, **16**:797-811.
224. Hitchcock CR, Sullivan WA, Wangenstein OH: **The value of achlorhydria as a screening test for gastric cancer: a 10-year report.** *Gastroenterology* 1955, **29**:621-628.
225. De Vita S, Sacco C, Sansonno D, Gloghini A, Dammacco F, Crovatto M, Santini G, Dolcetti R, Boiocchi M, Carbone A, Zagonel V: **Characterization of overt B-cell lymphomas in patients with hepatitis C.** *Blood* 1997, **90**:776-782.
226. Lipsmeyer EA: **Simultaneous development of autoimmune disease and malignancy in two elderly patients.** *J Am Geriatr Soc* 1979, **27**:455-458.

227. Sandilands GP, Gray A, Cooney A, Browning JD, Anderson JR: **Formation of auto-rosettes by peripheral blood lymphocytes.** *Clin Exp Immunol* 1975, **22**:493-501.
228. Paroli M, Barnaba V: **Mechanism of CD8+ T cell peripheral tolerance to our own antigens.** *Front Biosci* 2005, **10**:1628-1634.
229. Kessels HW, de Visser KE, Tirion FH, Coccoris M, Kruisbeek AM, Schumacher TN: **The impact of self-tolerance on the polyclonal CD8+ T cell repertoire.** *J Immunol* 2004, **172**:2324-2331.
230. Amedei A, Bergman MP, Appelmelk BJ, Azzurri A, Benaglio M, Tamburini C, van der Zee R, Telford JL, Vandembroucke-Grauls CMJE, D'Elios MM, Del Prete G: **Molecular mimicry between Helicobacter pylori antigens and H+, K+-adenosine triphosphatase in human gastric autoimmunity.** *J Exp Med* 2003, **198**:1147-1156.
231. Wucherpfennig KW, Strominger JL: **Molecular mimicry in T cell-mediated autoimmunity: viral peptides activate human T cell clones specific for myelin basic protein.** *Cell* 1995, **80**:695-705.
232. Hausmann S, Wucherpfennig KW: **Activation of autoreactive T cells by peptides from human pathogens.** *Curr Opin Immunol* 1997, **9**:831-838.
233. Voehringer D, Blaser C, Grawitz AB, Chisari FV, Buerki K, Pircher H: **Break of T cell ignorance to a viral antigen in the liver induces hepatitis.** *J Immunol* 2000, **165**:2415-2422.
234. Srinivasappa J, Saegusa J, Prabhakar BS, Gentry MK, Buchmeier MJ, Wiktor TJ, Koprowski H, Oldstone MB, Notkins AL: **Molecular mimicry: frequency of reactivity of monoclonal antiviral antibodies with normal tissues.** *J Virol* 1986, **57**:397-401.
235. Taurog JD, Richardson JA, Croft JT, Simmons WA, Zhou M, Fernandez-Sueiro JL, Balish E, Hammer RE: **The germfree state prevents development of gut and joint inflammatory disease in HLA-B27 transgenic rats.** *J Exp Med* 1994, **180**:2359-2364.
236. Fujinami RS, Oldstone MB: **Amino acid homology between the encephalitogenic site of myelin basic protein and virus: mechanism for autoimmunity.** *Science* 1985, **230**:1043-1045.
237. Singh VK, Nagaraju K: **Experimental autoimmune uveitis: molecular mimicry and oral tolerance.** *Immunol Res* 1996, **15**:323-346.
238. Garza KM, Tung KS: **Frequency of molecular mimicry among T cell peptides as the basis for autoimmune disease and autoantibody induction.** *J Immunol* 1995, **155**:5444-5448.
239. Ohashi PS, Oehen S, Buerki K, Pircher H, Ohashi CT, Odermatt B, Malissen B, Zinkernagel RM, Hengartner H: **Ablation of 'tolerance' and induction of diabetes by virus infection in viral antigen transgenic mice.** *Cell* 1991, **65**:305-317.
240. Oldstone MB, Nerenberg M, Southern P, Price J, Lewicki H: **Virus infection triggers insulin-dependent diabetes mellitus in a transgenic model: role of anti-self (virus) immune response.** *Cell* 1991, **65**:319-331.
241. Oldstone MB: **Molecular mimicry and immune mediated diseases.** *FASEB J* 1998, **12**:1255-1265.
242. Theofilopoulos AN, Kono DH: **Mechanisms and genetics of autoimmunity.** *Ann NY Acad Sci* 1998, **841**:225-235.
243. Albert LJ, Inman RD: **Molecular mimicry and autoimmunity.** *N Engl J Med* 1999, **341**:2068-2074.
244. Mamula MJ, Lin RH, Janeway CA Jr, Hardin JA: **Breaking T cell tolerance with foreign and self co-immunogens. A study of autoimmune B and T cell epitopes of cytochrome c.** *J Immunol* 1992, **149**:789-795.
245. Lin RH, Mamula MJ, Hardin JA, Janeway CA Jr: **Induction of autoreactive B cells allows priming of autoreactive T cells.** *J Exp Med* 1991, **173**:1433-1439.
246. Mamula MJ, Jemmerson R, Hardin JA: **The specificity of human anti-cytochrome c autoantibodies that arise in autoimmune disease.** *J Immunol* 1990, **144**:1835-1840.
247. Pircher H, Rohrer UH, Moskophidis D, Zinkernagel RM, Hengartner H: **Lower receptor avidity required for thymic clonal deletion than for effector T-cell function.** *Nature* 1991, **351**:482-485.
248. Sandberg JK, Franksson L, Sundback J, Michaelsson J, Petersson M, Achour A, Wallin RP, Sherman NE, Bergman T, Jornvall H, Hunt DF, Kiessling R, Karre K: **T cell tolerance based on avidity thresholds rather than complete deletion allows maintenance of maximal repertoire diversity.** *J Immunol* 2000, **165**:25-33.
249. Bouneaud C, Kourilsky P, Bousso P: **Impact of negative selection on the T cell repertoire reactive to a self-peptide: a large fraction of T cell clones escapes clonal deletion.** *Immunity* 2000, **13**:829-840.
250. Casanova JL, Cerottini JC, Matthes M, Necker A, Gournier H, Barra C, Widmann C, MacDonald HR, Lemonnier F, Malissen B, Mayanski JL: **H-2-restricted cytolytic T lymphocytes specific for HLA display T cell receptors of limited diversity.** *J Exp Med* 1992, **176**:439-447.
251. Kurts C, Sutherland RM, Davey G, Li M, Lew AM, Blanas E, Carbone FR, Miller JF, Heath WR: **CD8 T cell ignorance or tolerance to islet antigens depends on antigen dose.** *Proc Natl Acad Sci USA* 1999, **96**:12703-12707.
252. Thomis DC, Berg LJ: **The role of Jak3 in lymphoid development, activation, and signalling.** *Curr Opin Immunol* 1997, **9**:541-547.
253. Sadlack B, Kuhn R, Schorle H, Rajewsky K, Muller W, Horak I: **Development and proliferation of lymphocytes in mice deficient for both interleukins-2 and -4.** *Eur J Immunol* 1994, **24**:281-284.
254. Mier JW, Gallo RC: **Purification and some characteristics of human T-cell growth factor from phytohemagglutinin stimulated lymphocyte conditioned media.** *Proc Natl Acad Sci USA* 1980, **77**:6134-6138.
255. Griffin JD, Lowenberg B: **Clonogenic cells in acute myeloblastic leukemia.** *Blood* 1986, **68**:1185-1195.
256. Lotem J, Sachs L: **Hematopoietic cytokines inhibit apoptosis induced by transforming growth factor beta 1 and cancer chemotherapy compounds in myeloid leukemic cells.** *Blood* 1992, **80**:1750-1757.
257. Sachs L, Lotem J: **Control of programmed cell death in normal and leukemic cells: new implications for therapy.** *Blood* 1993, **82**:15-21.
258. Zubiaga AM, Munoz E, Huber BT: **IL-4 and IL-2 selectively rescue T cell subsets from glucocorticoid-induced apoptosis.** *J Immunol* 1992, **149**:107-112.
259. Hardin J, MacLeod S, Grigorieva I, Chang R, Barlogie B, Xiao H, Epstein J: **Interleukin-6 prevents dexamethasone-induced myeloma cell death.** *Blood* 1994, **84**:3063-3070.
260. Collins MK, Marvel J, Malde P, Lopez-Rivas A: **Interleukin 3 protects murine bone marrow cells from apoptosis induced by DNA damaging agents.** *J Exp Med* 1992, **176**:1043-1051.
261. Strasser A, Bouillet P: **The control of apoptosis in lymphocyte selection.** *Immunol Rev* 2003, **193**:82-92.
262. Greten FR, Eckmann L, Greten TF, Park JM, Li ZW, Egan LJ, Kagnoff MF, Karin M: **IKKbeta links inflammation and tumorigenesis in a mouse model of colitis-associated cancer.** *Cell* 2004, **118**(3):285-296.
263. Pikarsky E, Porat RM, Stein I, Abramovitch R, Amit S, Kasem S, Gutkovich-Pyest E, Urieli-Shoval S, Galun E, Ben-Neriah Y: **NF-kappaB functions as a tumour promoter in inflammation-associated cancer.** *Nature* 2004, **431**(7007):461-466.
264. Strasser A, Whittingham S, Vaux DL, Bath ML, Adams JM, Cory S, Harris AW: **Enforced BCL2 expression in B-lymphoid cells prolongs antibody responses and elicits autoimmune disease.** *Proc Natl Acad Sci USA* 1991, **88**:8661-8665.
265. Watanabe-Fukunaga R, Brannan CI, Copeland NG, Jenkins NA, Nagata S: **Lymphoproliferation disorder in mice explained by defects in Fas antigen that mediates apoptosis.** *Nature* 1992, **356**:314-317.
266. Grodzicky T, Elkon KB: **Apoptosis: A case where too much or too little can lead to autoimmunity.** *Mount Sinai J Med* 2002, **69**:208-219.
267. Sachs L: **The control of hematopoiesis and leukemia: From basic biology to the clinic.** *Proc Natl Acad Sci USA* 1996, **93**:4742-4749.
268. Strasser A, Harris AW, Bath ML, Cory S: **Novel primitive lymphoid tumours induced in transgenic mice by cooperation between myc and bcl-2.** *Nature* 1990, **348**:331-333.
269. de Thonel A, Eriksson JE: **Regulation of death receptors-Relevance in cancer therapies.** *Toxicol Appl Pharmacol* 2005, **207**(2 Suppl):123-132.
270. Drappa J, Vaishnav AK, Sullivan KE, Chu JL, Elkon KB: **Fas gene mutations in the Canale-Smith syndrome, an inherited lymphoproliferative disorder associated with autoimmunity.** *N Engl J Med* 1996, **335**:1643-1649.
271. Fisher GH, Rosenberg FJ, Straus SE, Dale JK, Middleton LA, Lin AY, Strober W, Lenardo MJ, Puck JM: **Dominant interfering Fas gene mutations impair apoptosis in a human autoimmune lymphoproliferative syndrome.** *Cell* 1995, **81**:935-946.

272. O'Connell J, Bennett MW, O'Sullivan GC, O'Callaghan J, Collins JK, Shanahan F: **Expression of Fas (CD95/APO-1) ligand by human breast cancers: significance for tumor immune privilege.** *Clin Diagn Lab Immunol* 1999, **6**:457-463.
273. Bodey B, Bodey B Jr, Siegel SE, Kaiser HE: **Immunocytochemical detection of leukocyte-associated and apoptosis-related antigen expression in childhood brain tumors.** *Crit Rev Oncol Hematol* 2001, **39**:3-16.
274. Saas P, Walker PR, Hahne M, Quiquerez AL, Schnuriger V, Perrin G, French L, Van Meir EG, de Tribolet N, Tschopp JJ, Dietrich PY: **Fas ligand expression by astrocytoma in vivo: maintaining immune privilege in the brain.** *J Clin Invest* 1997, **99**:1173-1178.
275. Mitsiades N, Poulaki V, Kotoula V, Leone A, Tsokos M: **Fas ligand is present in tumors of the Ewing's sarcoma family and is cleaved into a soluble form by a metalloproteinase.** *Am J Pathol* 1998, **153**:1947-1956.
276. Koyama S, Koike N, Adachi S: **Fas receptor counterattack against tumor-infiltrating lymphocytes in vivo as a mechanism of immune escape in gastric carcinoma.** *J Cancer Res Clin Oncol* 2001, **127**:20-26.
277. Que FG, Phan VA, Phan VH, Celli A, Batts K, LaRusso NF, Gores GJ: **Cholangiocarcinomas express Fas ligand and disable the Fas receptor.** *Hepatology* 1999, **30**:1398-1404.
278. Tinhofer I, Marschitz I, Kos M, Henn T, Egle A, Villunger A, Greil R: **Differential sensitivity of CD4+ and CD8+ T lymphocytes to the killing efficacy of Fas (Apo-1/CD95) ligand+ tumor cells in B chronic lymphocytic leukemia.** *Blood* 1998, **91**:4273-4281.
279. O'Connell J, Bennett MW, O'Sullivan GC, Roche D, Kelly J, Collins JK, Shanahan F: **Fas ligand expression in primary colon adenocarcinomas: evidence that the Fas counterattack is a prevalent mechanism of immune evasion in human colon cancer.** *J Pathol* 1998, **186**:240-246.
280. Shiraki K, Tsuji N, Shiroda T, Isselbacher KJ, Takahashi H: **Expression of Fas ligand in liver metastases of human colonic adenocarcinomas.** *Proc Natl Acad Sci USA* 1997, **94**:6420-6425.
281. O'Connell J, O'Sullivan GC, Collins JK, Shanahan F: **The Fas counterattack: Fas-mediated T cell killing by colon cancer cells expressing Fas ligand.** *J Exp Med* 1996, **184**:1075-1082.
282. Young MR, Wright MA, Lozano Y, Mathews JP, Benefield J, Prechel MM: **Mechanisms of immune suppression in patients with head and neck cancer: influence on the immune infiltrate of the cancer.** *Int J Cancer* 1996, **67**:333-338.
283. Niehans GA, Brunner T, Frizelle SP, Liston JC, Salerno CT, Knapp DJ, Green DR, Kratzke RA: **Human lung carcinomas express Fas ligand.** *Cancer Res* 1997, **57**:1007-1012.
284. Bennett MW, O'Connell J, O'Sullivan GC, Brady C, Roche D, Collins JK, Shanahan F: **The Fas counterattack in vivo: apoptotic depletion of tumor infiltrating lymphocytes associated with Fas ligand expression by human esophageal carcinoma.** *J Immunol* 1998, **160**:5669-5675.
285. Rabinowich H, Reichert TE, Kashii Y, Gastman BT, Bell MC, Whiteside TL: **Lymphocyte apoptosis induced by Fas ligand-expressing ovarian carcinoma cells: implications for altered expression of TCR in tumor.** *J Clin Invest* 1998, **101**:2579-2588.
286. Sinkovics JG: **Malignant lymphoma arising from natural killer cells: report of the first case in 1970 and newer developments in the FasL-->FasR system.** *Acta Microbiol Immunol Hung* 1997, **44**:295-303.
287. von Bernstorff W, Spanjaard RA, Chan AK, Lockhart DC, Sadanaga N, Wood I, Peiper M, Goedegebuure PS, Eberlein TJ: **Pancreatic cancer cells can evade immune surveillance via nonfunctional Fas(APO-1/CD95) receptors and aberrant expression of functional Fas ligand.** *Surgery* 1999, **125**:73-84.
288. Hahne M, Rimoldi D, Schroter M, Romero P, Schreiber M, French LE, Schneider P, Bornand T, Fontana A, Lienard D, Cerottini J, Tschopp J: **Melanoma cell expression of Fas(Apo-1/CD95) ligand: implications for tumor immune escape.** *Science* 1996, **274**:1363-1366.
289. Sinkovics JG, Horvath JC: **Virological and immunological connotations of apoptotic and anti-apoptotic forces in neoplasia.** *Int J Oncol* 2001, **19**:473-488.
290. Graban J, Kohut A: **Apoptosis in T-lymphocytes and its significance.** *Cesk Fysiol* 2003, **52**:144-52.
291. Strand S, Hofmann WJ, Hug H, Muller M, Otto G, Strand D, Mariani SM, Stremmel W, Krammer PH, Galle PR: **Lymphocyte apoptosis induced by CD95 (APO-1/Fas) ligand-expressing tumor cells - a mechanism of immune evasion?** *Nat Med* 1996, **2**:1361-1366.
292. Chappell DB, Restifo NP: **T cell-tumor cell: a fatal interaction?** *Cancer Immunol Immunother* 1998, **47**:65-71.
293. Tyler KL, Fields BN: **Pathogenesis of viral infections.** In *Fields Virology* Edited by: Fields BN, Knipe DM, Howley PM. Philadelphia, Lippincott-Raven Publishers; 1996:173-203.
294. Citovsky V, Rottem S, Nussbaum O, Laster Y, Rott R, Loyter A: **Animal viruses are able to fuse with prokaryotic cells. Fusion between Sendai or influenza virions and Mycoplasma.** *J Biol Chem* 1988, **263**:461-467.
295. Ketcham CM, Anai S, Reutzel R, Sheng S, Schuster SM, Brenes RB, Agbandje-McKenna M, McKenna R, Rosser CJ, Boehlein SK: **p37 Induces tumor invasiveness.** *Mol Cancer Ther* 2005, **4**(7):1031-1038.
296. Lawrence JG: **Horizontal and vertical gene transfer: the life history of pathogens.** *Contrib Microbiol* 2005, **12**:255-271.
297. Holmgren L, Szeles A, Rajnavolgyi E, Folkman J, Klein G, Ernberg I, Falk KI: **Horizontal transfer of DNA by the uptake of apoptotic bodies.** *Blood* 1999, **93**(11):3956-3963.
298. Spetz AL, Patterson BK, Lore K, Andersson J, Holmgren L: **Functional gene transfer of HIV DNA by an HIV receptor-independent mechanism.** *J Immunol* 1999, **163**(2):736-742.
299. Sloots A, Wels WS: **Recombinant derivatives of the human high-mobility group protein HMGB2 mediate efficient non-viral gene delivery.** *FEBS J* 2005, **272**(16):4221-4236.
300. Houghton J, Stoicov C, Nomura S, Rogers AB, Carlson J, Li H, Cai X, Fox JG, Goldenring JR, Wang TC: **Gastric cancer originating from bone marrow-derived cells.** *Science* 2004, **306**(5701):1568-1571.
301. Contardi E, Palmisano GL, Tazzari PL, Martelli AM, Fala F, Fabbi M, Kato T, Lucarelli E, Donati D, Polito L, Bolognesi A, Ricci F, Salvi S, Gargaglione V, Mantero S, Alberghini M, Ferrara GB, Pistillo MP: **CTLA-4 is constitutively expressed on tumor cells and can trigger apoptosis upon ligand interaction.** *Int J Cancer* 2005, **117**(4):538-550.
302. Carrel S, De Tribolet N, Gross N: **Expression of HLA-DR and common acute lymphoblastic leukemia antigens on glioma cells.** *Eur J Immunol* 1982, **12**(4):354-357.
303. Carrel S, Schmidt-Kessen A, Mach JP, Heumann D, Girardet C: **Expression of common acute lymphoblastic leukemia antigen (CALLA) on human malignant melanoma cell lines.** *J Immunol* 1983, **130**(5):2456-2460.
304. Paydas S, Tanriverdi K, Yavuz S, Disel U, Baslamisli F, Burgut R: **PRAME mRNA levels in cases with acute leukemia: clinical importance and future prospects.** *Am J Hematol* 2005, **79**(4):257-261.
305. Dahlke MH, Larsen SR, Rasko JE, Schlitt HJ: **The biology of CD45 and its use as a therapeutic target.** *Leuk Lymphoma* 2004, **45**(2):229-236.
306. Warnke RA, Rouse RV: **Limitations encountered in the application of tissue section immunodiagnosis to the study of lymphomas and related disorders.** *Hum Pathol* 1985, **16**:326-331.
307. McDonnell JM, Beschoner WE, Kuhajda FP, Dement SH: **Common leukocyte antigen staining sarcoma.** *Cancer* 1987, **59**:1438-1441.
308. Nandedkar MA, Palazzo J, Abbondanzo SL, Lasota J, Miettinen M: **CD45 (leukocyte common antigen) immunoreactivity in metastatic undifferentiated and neuroendocrine carcinoma: a potential diagnostic pitfall.** *Mod Pathol* 1998, **11**(12):1204-1210.
309. Ruff MR, Pert CB: **Small cell carcinoma of the lung: macrophage-specific antigens suggest hemopoietic stem cell origin.** *Science* 1984, **225**:1034-1036.
310. Ball ED, Sorensen GD, Pettengill OS: **Expression of myeloid and major histocompatibility antigens on small cell carcinoma of the lung cell lines analyzed by cytofluorography: modulation by γ -interferon.** *Cancer Res* 1986, **46**:2335-2339.
311. Lipinski M, Braham K, Caillaud J-M, Tursz T: **HNK-1 antibody detects an antigen expressed on neuroectodermal cells.** *J Exp Med* 1983, **158**:1775-1780.
312. Papadimitriou CS, Bai MK, Kotsiantzi AJ, Costopoulos JS, Hytioglou P: **Phenotype of Hodgkin and Sternberg-Reed cells and expression of CD57 (LEU7) antigen.** *Leuk Lymphoma* 1995, **20**(1-2):125-130.

313. Stoler DL, Chen N, Basik M, Kahlenberg MS, Rodriguez-Bigas MA, Petrelli NJ, Anderson GR: **The onset and extent of genomic instability in sporadic colorectal tumor progression.** *Proc Natl Acad Sci USA* 1999, **96**:15121-15126.
314. Degos L: **All-trans-retinoic acid treatment and retinoic acid receptor alpha gene rearrangement in acute promyelocytic leukemia: a model for differentiation therapy.** *Int J Cell Cloning* 1992, **10**:63-69.
315. Griffiths JD, McKinna JA, Rowbotham HD, Tsolakidis P, Salsbury AJ: **Carcinoma of the colon and rectum: circulating malignant cells and five-year survival.** *Cancer* 1973, **31**:226-236.
316. Koo J, Fung K, Siu KF, Lee NW, Lett Z, Ho J, Wong J, Ong GB: **Recovery of malignant tumor cells from the right atrium during hepatic resection for hepatocellular carcinoma.** *Cancer* 1983, **52**:1952-1956.
317. Sako K, Marchetta FC: **Radioautography of in vitro labeled tumor cells in postoperative wound drainage.** *Cancer* 1966, **19**:735-737.
318. Molnar B, Ladanyi A, Tanko L, Sreter L, Tulassay Zs: **Circulating Tumor Cell Clusters in the Peripheral Blood of Colorectal Cancer Patients.** *Clin Cancer Res* 2001, **7**:4080-4085.
319. Liotta LA, Kleinerman J, Sidel GM: **The significance of hematogenous tumor cell clumps in the metastatic process.** *Cancer Res* 1976, **36**:889-894.
320. Tirapu I, Mazzolini G, Rodriguez-Calvillo M, Arina A, Palencia B, Gabari I, Melero I: **Effective Tumor Immunotherapy: Start the engine, release the brakes, step on the gas pedal, ... and get ready to face autoimmunity.** *Arch Immunol Ther Exp* 2002, **50**:13-18.
321. Smith CB, Friedewald WT, Chanock RM: **Shedding of Mycoplasma pneumoniae after tetracycline and erythromycin therapy.** *N Engl J Med* 1967, **276**:1172-1175.
322. Szathmary S: **Immunomodulation of pathogen-host interactions.** In *PhD Thesis* Szent Istvan University, Faculty of Veterinary Medicine, Budapest Hungary; 2005.
323. Otton R, Curi R: **Toxicity of a mixture of fatty acids on human blood lymphocytes and leukaemia cell lines.** *Toxicol In Vitro* 2005, **19**:749-755.
324. Kovacs A: *Personal communication* .
325. Hetzel BS, Clugston GA: **Iodine.** In *Nutrition in Health and Disease Volume 9*. Edited by: Shils M, Olson JA, Shike M, Ross AC. Baltimore. Williams & Wilkins; 1999:253-264.
326. Benmiloud M, Chaouki ML, Gutekunst R, Teichert HM, Wood WG, Dunn JT: **Oral iodized oil for correcting iodine deficiency: optimal dosing and outcome indicator selection.** *J Clin Endocrinol Metab* 1994, **79**:20-24.
327. Abuye C, Hailemariam B, Tibeb HN, Urga K, Gebru H: **The effect of varying doses of oral iodized oil in the prophylaxis of endemic goitre in elementary schools children.** *Ethiop Med J* 1995, **33**:115-123.
328. Heimburger DC, Alexander CB, Birch R, Butterworth CE Jr, Bailey WC, Krumdieck CL: **Improvement in bronchial squamous cell metaplasia in smokers treated with folic acid and vitamin B-12. Report of a preliminary randomized, double-blind intervention trial.** *JAMA* 1988, **259**:1525-1530.
329. Saito M, Kato H, Tsuchida T, Konaka C: **Chemoprevention effects on bronchial squamous metaplasia by folate and vitamin B-12.** *Chest* 1994, **106**:496-499.
330. Gonzalez MJ, Miranda-Massari JR, Mora EM, Guzman A, Riordan NH, Riordan HD, Casciari JJ, Jackson JA, Roman-Franco A: **Ascorbic acid and cancer 25 years later.** *Integrative Cancer Ther* 2005, **4**:35-44.
331. Worthington V: **Analyzing data to compare nutrients in conventional versus organic crops.** *J Alternat Complement Med* 2002, **8**:529-532.
332. Strickler HD, Wylie-Rosett J, Rohan T, Hoover DR, Smoller S, Burk RD, Yu H: **The relation of type 2 diabetes and cancer.** *Diabetes Technol Ther* 2001, **3**:263-274.
333. Frank BH, Manson JE, Liu S, Hunter D, Colditz GA, Michels KB, Speizer FE, Giovannucci E: **Prospective study of adult onset diabetes mellitus (Type 2) and risk of colorectal cancer in women.** *J Natl Cancer Inst* 1999, **91**:542-547.

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